

850 Professional IC



Anion – MSM-HC – MCS – Gradient – 2.850.2240

Manual
8.850.8027EN



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2.850.2240**

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1 Introduction

1.1 Instrument description

The instrument **850 Professional IC – Anion – MSM-HC – MCS – Gradient** (2.850.2240) is one of the model versions of the Professional IC line of instruments manufactured by the Metrohm Company. The Professional IC line of instruments is distinguished by:

- the **intelligence** of its components, which are able to monitor and optimize all functions and to provide documentation according to FDA requirements.
- its **compactness**.
- its **flexibility**. A suitable model version exists for every application. Individual instruments can be converted, expanded or modified to create a different instrument as needed.
- its **transparency**. All components are easily accessible and arranged in a clear manner.
- its **safety**. Chemicals and electronics are separated and a leak sensor is integrated in the wet end.
- its **environmental compatibility**.
- **low noise emission**.

The instrument is operated with **MagIC Net** software. It is connected via a USB connection to a PC on which MagIC Net is installed. The software automatically recognizes the instrument and checks its functional readiness. MagIC Net controls and monitors the instrument, evaluates the measured data and administers it in a database. The operation of MagIC Net is described in the online help or in the tutorial for MagIC Net.

The low pressure mixing valve and two additional eluent degassers in the lower area of the instrument are used for the controlled mixing of up to 3 eluents.

The instrument contains the following components:

Eluent degasser

The eluent degasser removes gas bubbles and dissolved gases from the eluent. For degassing, the eluent flows into a vacuum chamber through a special fluoropolymer capillary.

High pressure pump

The intelligent and low pulsation high pressure pump pumps the eluent through the system. It is equipped with a chip on which its technical specifications and "life history" (operating hours, service data, ...) are saved.



Inline filter

Inline filters protect the separation column securely against possible contamination from the eluent. Inline filters can however also just as well be used for the purpose of protecting other sensitive components against contaminations in the solutions used. The fine 2 µm material of the readily and easily replaceable filter platelets removes particles such as bacteria and algae from the solutions.

Pulsation damper

The pulsation damper protects the separation column from damage caused by pressure fluctuations when switching the injection valve, and reduces interfering pulsations during highly sensitive measurements.

Sample degasser

The sample degasser removes gas bubbles and dissolved gases from the sample. For degassing, the sample flows into a vacuum chamber through a special fluoropolymer capillary.

Injection valve

The injection valve connects the eluent and sample path through rapid and precise valve switchover. A precisely measured amount of sample solution is injected and rinsed with eluent onto the separation column.

Column thermostat

The column thermostat controls the temperature of the column and eluent channel and thus ensures stable measuring conditions. It provides space for 2 separation columns.

High Capacity Metrohm Suppressor Module (MSM-HC)

The MSM-HC is used for chemical suppression with high capacity during anion analysis. It is pressure-stable, robust and resistant to solvents.

Peristaltic pump

The Peristaltic pump is used for pumping sample and auxiliary solutions. It can rotate in both directions.

Metrohm CO₂ suppressor (MCS)

The MCS removes the CO₂ from the eluent flow. This reduces the background conductivity, improves the detection sensitivity and minimizes the injection and carbonate peaks.

Conductivity detector

The conductivity detector continuously measures the conductivity of the liquid passing through and indicates these signals in digital form (DSP – Digital Signal Processing). The conductivity detector exhibits outstanding thermal stability and thus guarantees reproducible measuring conditions.

Separation column

The intelligent separation column is the heart of the ion chromatographic analysis. It separates the different components corresponding to their interactions with the column. The Metrohm analytical columns are equipped with a chip on which their technical specifications and their history (first use / setting up, operating hours, injections, ...) are saved.

1.2 Intended use

The **850 Professional IC – Anion – MSM-HC – MCS – Gradient** is used for the ion chromatography determination of anions with binary **high pressure gradient** and **sequential suppression**:

Sequential suppression consists of:

- Chemical suppression by the Metrohm Suppressor Module (MSM) and subsequent
- CO₂ suppression by the Metrohm CO₂ Suppressor (MCS) (*see Chapter 3.18, page 60*).

The use of this technology reduces background conductivity to an absolute minimum.

If required, the instrument can also be used for the determination of cations and anions without suppression.

The present instrument is suitable for processing chemicals and flammable samples. The usage of the 850 Professional IC – Anion – MSM-HC – MCS – Gradient therefore requires that the user has basic knowledge and experience in the handling of toxic and caustic substances. Knowledge with respect to the application of the fire prevention measures prescribed for laboratories is also mandatory.

1.3 About the documentation



Caution

Please read through this documentation carefully before putting the instrument into operation. The documentation contains information and warnings which have to be followed by the user in order to ensure safe operation of the instrument.

1.3.1 Symbols and conventions

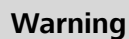
The following symbols and styles are used in this documentation:

(5-12)	Cross-reference to figure legend The first number refers to the figure number, the second to the instrument part in the figure.
1	Instruction step Carry out these steps in the sequence shown.
	Warning This symbol draws attention to a possible life hazard or risk of injury.
	Warning This symbol draws attention to a possible hazard due to electrical current.
	Warning This symbol draws attention to a possible hazard due to heat or hot instrument parts.
	Warning This symbol draws attention to a possible biological hazard.
	Caution This symbol draws attention to a possible damage of instruments or instrument parts.
	Note This symbol marks additional information and tips.

1.4.1 General notes on safety



This instrument has left the factory in a flawless state in terms of technical safety. To maintain this state and ensure non-hazardous operation of the instrument, the following instructions must be observed carefully.



Only operate this instrument with a mains voltage specified for it (see rear panel of the instrument).



Protection against electrostatic charges



Warning

Electronic components are sensitive to electrostatic charges and can be destroyed by discharges.

Always pull the mains cable out of the mains connection socket before connecting or disconnecting electrical appliances on the rear panel of the instrument.

1.4.3 Tubing and capillary connections



Caution

Leaks in tubing and capillary connections are a safety risk. Tighten all connections well by hand. Avoid applying excessive force to tubing connections. Damaged tubing ends lead to leakage. Appropriate tools can be used to loosen connections.

Check the connections regularly for leakage. If the instrument is used mainly in unattended operation, then weekly inspections are mandatory.

1.4.4 Flammable solvents and chemicals

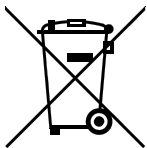


Warning

All relevant safety measures are to be observed when working with flammable solvents and chemicals.

- Set up the instrument in a well-ventilated location (e.g. laboratory flue).
- Keep all sources of flame far from the workplace.
- Clean up spilled fluids and solids immediately.
- Follow the safety instructions of the chemical manufacturer.

1.4.5 Recycling and disposal



This product is covered by European Directive 2002/96/EC, WEEE – Waste from Electrical and Electronic Equipment.

The correct disposal of your old equipment will help to prevent negative effects on the environment and public health.

More details about the disposal of your old equipment can be obtained from your local authorities, from waste disposal companies or from your local dealer.

2 Overview of the instrument

2.1 Front

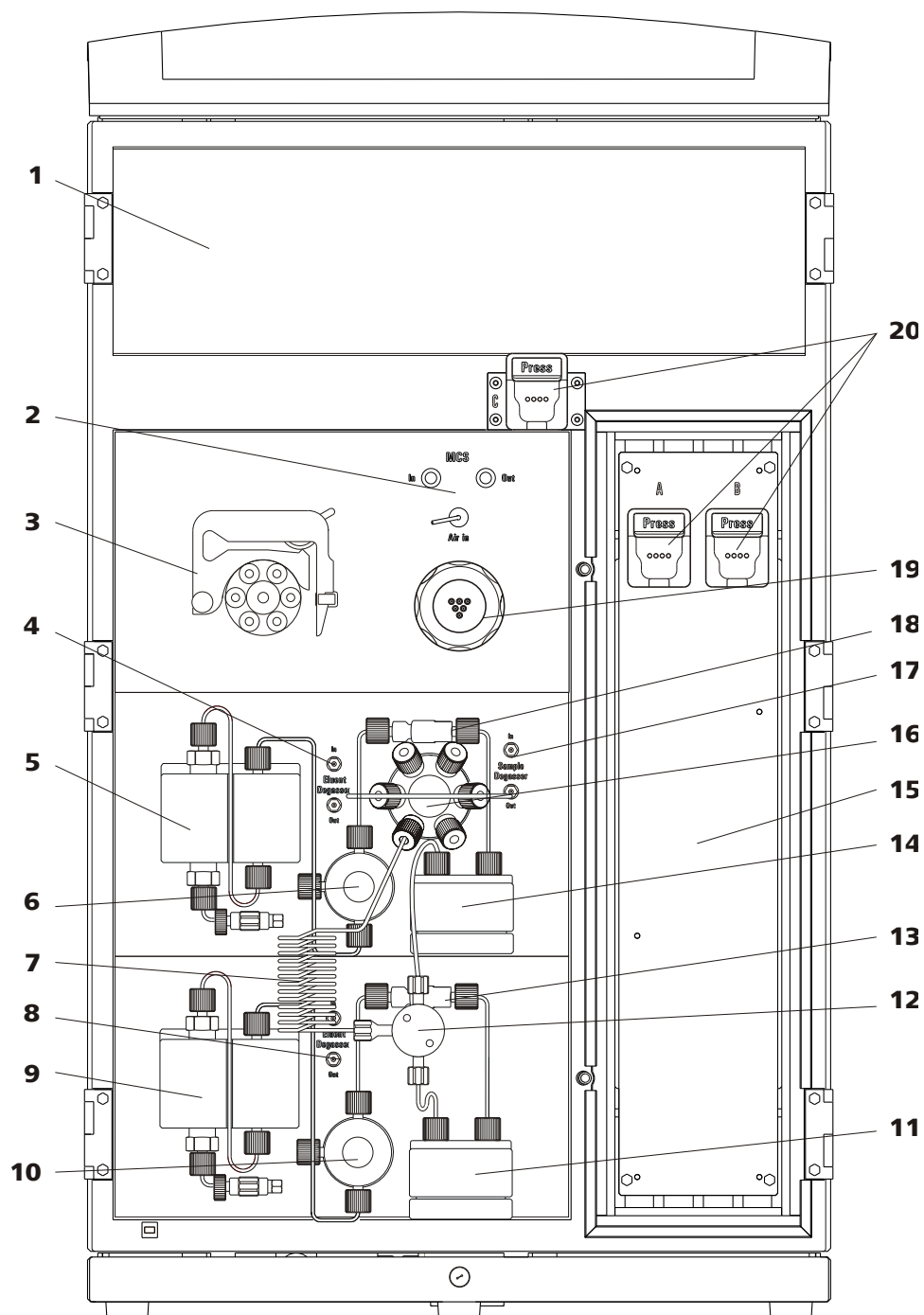


Figure 1 Front 850 Professional IC – Anion – MSM-HC – MCS – Gradient

- ## 1 Detector chamber

Space for the conductivity detector (see Chapter 3.19, page 63).

- ## 2 MCS

See Chapter 3.18, Page 60.



Caution

When connecting an instrument to the MSB connector (2-**6**) you **must** switch off the 850 Professional IC .

3 Installation

3.1 About this chapter

The *Installation* chapter contains

- this overview
- a brief set of instructions for the initial installation of the 850 Professional IC – Anion – MSM-HC – MCS – Gradient (see *Chapter 3.2, page 13*) as an isocratic system. At each step you will find cross-references to more detailed installation instructions for individual components, should you require such aids.
- an installation diagram (see *Figure 3, page 17*), showing a completely installed instrument.
- several chapters with detailed installation instructions for all components, including those that are already installed at the time the instrument is delivered.

3.2 Initial installation



Note

A large number of the capillary connections are already connected at the time the instrument is delivered.

Installing 850 Professional IC – Anion – MSM-HC – MCS – Gradient

1 Setting up the instrument

See *Chapter 3.4, Page 19*.

2 Installations on the rear of the instrument

- Remove handle and rollers (see *Chapter 3.6.1, page 21*).
- Place the detector in the instrument and connect it (see *Chapter 3.6.2, page 24*).
- Remove transport locking screws (see *"Removing transport locking screws", page 26*).
- Connect the leak sensor (see *"Connecting the leak sensor", page 26*).



- Connect drainage tubings (see "Installing drainage tubing", page 29).

3 Connecting the eluent path

- Equip an eluent aspiration tubing for each of the two high pressure pumps and connect with the eluent bottle (*see Chapter 3.8.1, page 32*).
- Instead of the column, connect the 6.2744.040 coupling with a 6.2744.010 pressure screw at the end of the pre-installed column inlet capillary.
- Use a 6.2744.014 pressure screw to connect the capillary of the MSM-HC labeled with *Eluent* to the other end of the 6.2744.040 coupling.
- Use a 6.2744.090 long pressure screw to connect the capillary of the MSM-HC labeled with *Detector* to the input of the MCS (*see Chapter 3.18.2, page 60*).
- Connect the detector inlet capillary with a 6.2744.090 long pressure screw to the output of the MCS (*see Chapter 3.18.2, page 60*).

4 Connecting the sample path



Note

The sample degasser does not have to be connected. We recommend the usage of the sample degasser only if the sample matrix requires it.

- Connect the 6.1803.040 sample aspirating capillary connected at the sample input of the injection valve to the output of the sample degasser with the aid of a 6.2744.090 long pressure screw (see Chapter 3.13, page 45).
- Connect a part of the 6.1803.040 PTFE capillary with a 6.2744.090 long pressure screw to the input of the sample degasser. Guide the other end out of the instrument through a capillary feed-through and connect it to the Sample Processor.
- Guide the sample outlet capillary connected to the sample output of the injection valve out of the instrument through a capillary feed-through and onward to the waste container and then fasten it there.

5 Installing the peristaltic pump

(see Chapter 3.17.2, page 56).

- Connect the 6.1803.020 aspirating capillary for regeneration solutions with a 6.2744.034 tubing olive and a 6.2744.070 short pressure screw to the aspiration end of the 6.1826.320 pump tubing for the regeneration solution.
- Place the pump tubing in a tubing cartridge.
- Connect the 6.1803.020 aspirating capillary for rinsing solutions with a 6.2744.034 tubing olive and a 6.2744.070 short pressure screw to the aspiration end of the second 6.1826.320 pump tubing for the rinsing solution.
- Place the second pump tubing in the second tubing cartridge.
- Place both tubing cartridges into the peristaltic pump.

6 Connecting the MSM-HC

(see Chapter 3.16.2, page 52)

- Connect the capillary labeled with *H2SO4* with a 6.2744.180 pump tubing connection and a 6.2744.070 short pressure screw to the peristaltic pump on the outlet end of the pump tubing for the regeneration solution.
- Connect the capillary labeled with *H2O* with a 6.2744.180 pump tubing connection and a 6.2744.070 short pressure screw to the peristaltic pump on the outlet end of the pump tubing for the rinsing solution.
- Guide the two capillaries labeled with *Waste* into a waste container and fasten them there.

7 Connecting the instrument

- Connecting the instrument to the PC with a 6.2151.020 USB cable (see Chapter 3.20.1, page 66).
- Connect instrument to mains supply (see Chapter 3.20, page 66).

8 Initial start-up

(see Chapter 4.1, page 71)

- Switch on the PC and start MagIC Net.
- Switch on the instrument.
- Deaerate the high pressure pump (see Chapter 3.10.2, page 40).
- Set contact pressure of the peristaltic pump (see "Set flow rate", page 59).
- Rinse instrument without column.

9 Installing guard and separation column

- Remove the 6.2744.040 coupling between the column inlet capillary and the eluent inlet capillary of the MSM-HC.
- Connect guard column (optional) (*see Chapter 3.21, page 67*)
 - Fasten the guard column to the end of the column inlet capillary according to the specifications contained in the leaflet accompanying the guard column.
 - Rinse guard column.
- Connect the separation column (*see Chapter 3.22, page 69*)
 - Fasten the inlet of the separation column either to the end of the column inlet capillary or to the guard column (if used) according to the specifications contained in the leaflet accompanying the column.
 - Fasten the MSM-HC capillary labeled with *Eluent* with a 6.2744.070 PEEK pressure screw at the output of the separation column.
- Hang separation column with chip in the column holder of the instrument.

10 Conditioning the instrument

(see "Conditioning the system", page 72)

3.3 Installation diagram

Figure 3 Installation diagram shows the capillary connections with the application of the gradient technique and sequential suppression (MSM-HC and MCS).

The arrangement of the modules in the diagram corresponds to the front view of the instrument. Liquid containers (eluent bottle, sample vessel, waste container, auxiliary solutions container) and guard column (see *Chapter 3.21, page 67*) are not shown in the diagram.

Most of the capillaries are already pre-installed at the time the instrument is delivered. Capillaries on which nothing needs to be done at the time of initial installation are not numbered in the diagram.



2 Column inlet capillary

Connected to the injection valve and threaded into the capillary recesses of the column thermostat.

3	Connection capillary Capillary of the MSM-HC labeled with <i>Eluent</i> .
5	Detector inlet capillary
7	PTFE connection capillary Section of the 6.1803.040 PTFE capillary. Connects sample degasser with sample processor (required only if the sample degasser is to be used).
9	Rinsing solution aspirating capillary 6.1803.020 Connection to the bottle with rinsing solution.
11	Connection capillary for the rinsing solution Capillary of the MSM-HC labeled with <i>H2O</i> .
13	Regeneration solution aspirating capillary 6.1803.020 Connection to the regeneration solution bottle.
15	Connection capillary for the regeneration solution Capillary of the MSM-HC labeled with <i>H2SO4</i> .
17	Air aspirating capillary For aspirating CO ₂ -low air from the cartridges of the MCS.
19	PEEK pressure screws, long 6.2744.090
21	Pump tubing connection with safety device 6.2744.180
4	Connection capillary Capillary of the MSM-HC labeled with <i>Detector</i> .
6	Sample aspirating capillary 6.1803.040 Connected to the injection valve. The other end can be connected either to the sample degasser or guided directly out of the instrument.
8	Sample outlet capillary Connected to the injection valve.
10	Pump tubing 6.1826.320 Pump tubing with yellow-orange stoppers for conveying the rinsing solution.
12	Rinsing solution outlet capillary Capillary of the MSM-HC labeled with <i>Waste</i> .
14	Pump tubing 6.1826.320 Pump tubing with yellow-orange stoppers for conveying the regeneration solution.
16	Regeneration solution outlet capillary Capillary of the MSM-HC labeled with <i>Waste</i> .
18	PEEK pressure screws, short 6.2744.070
20	Tubing olive 6.2744.030

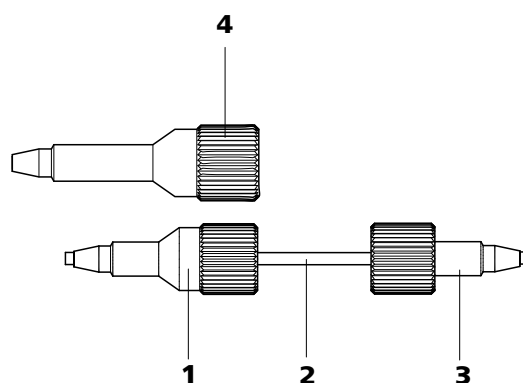


Figure 4 Connection of capillaries with pressure screws

1 PEEK pressure screw 6.2744.014 Use on the injection valve.	2 Connection capillary
3 PEEK pressure screw, short 6.2744.070 For use on the high pressure pump, the purge valve, the inline filter, the pulsation damper, the guard column and the separation column.	4 PEEK pressure screw, long 6.2744.090 Usage on other components. Is not used on all instruments.

Note

In order to keep the dead volume as low as possible, capillary connections should generally be as short as possible.

Note

For an improved overview, capillary and tubing connections can be bundled with the 6.1815.010 spiral band.

Connection capillary

PEEK capillaries and PTFE capillaries are used in the IC system.

PEEK capillaries (poly-
etheretherketone)

PEEK capillaries are temperature-resistant up to 100°C, stable under pressure up to 400 bar, flexible, chemically inert and exhibit an extremely smooth surface. They can be readily cut down to the desired length with the capillary cutter.

Usage:

- 6.1831.010 PEEK capillary (internal diameter of 0.25 mm) for the entire high pressure range.
- 6.1831.030 PEEK capillary (internal diameter of 0.75 mm) for sample handling in the ultra trace range.



PTFE capillaries (polytetrafluoroethylene)

Usage:

- PTFE capillaries with internal diameter of 0.5 mm for sample handling.
- PTFE capillaries with internal diameter of 0.97 mm for the sample handling as for rinsing and regeneration solutions (they do not have to be in the scope of delivery of the instrument).



3.6 Rear of the instrument

3.6.1 Rollers and handle

In order to make transport easier, the instrument is equipped with rollers and a handle.

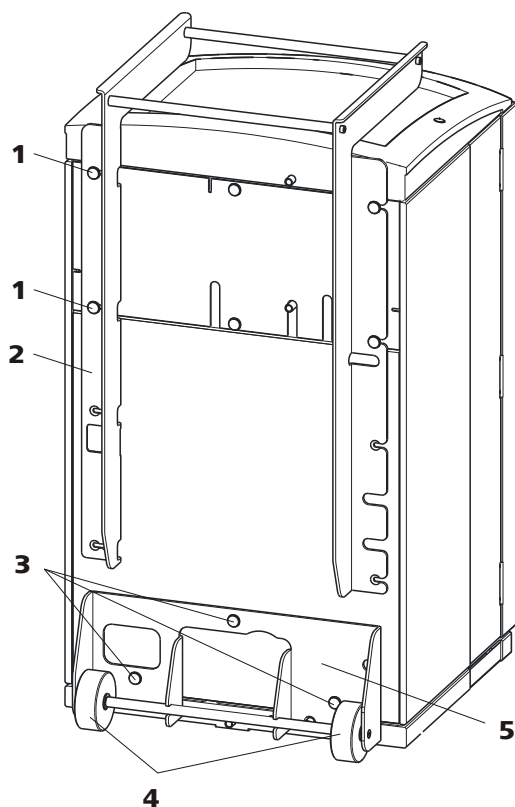


Figure 5 Rollers and handle

- | | |
|--|-------------------------|
| <p>1 Knurled screws</p> <p>For fastening the handle (5-2) and the rear panel of the detector chamber.</p> | <p>2 Handle</p> |
| <p>3 Knurled screws</p> <p>For fastening the roller holder (5-5).</p> | <p>4 Rollers</p> |
| <p>5 Roller holder</p> | |

Removing handle

- 1** Loosen knurled screws (5-**1**) and remove handle (5-**2**).

Removing rollers

Proceed as follows to remove the rollers:

- 1 Remove knurled screws (5-3).
- 2 Remove roller holder (5-5).

Mounting handle as MPak holder



- 1** Move handle (6-**2**) upwards and screw in the knurled screws (6-**1**)

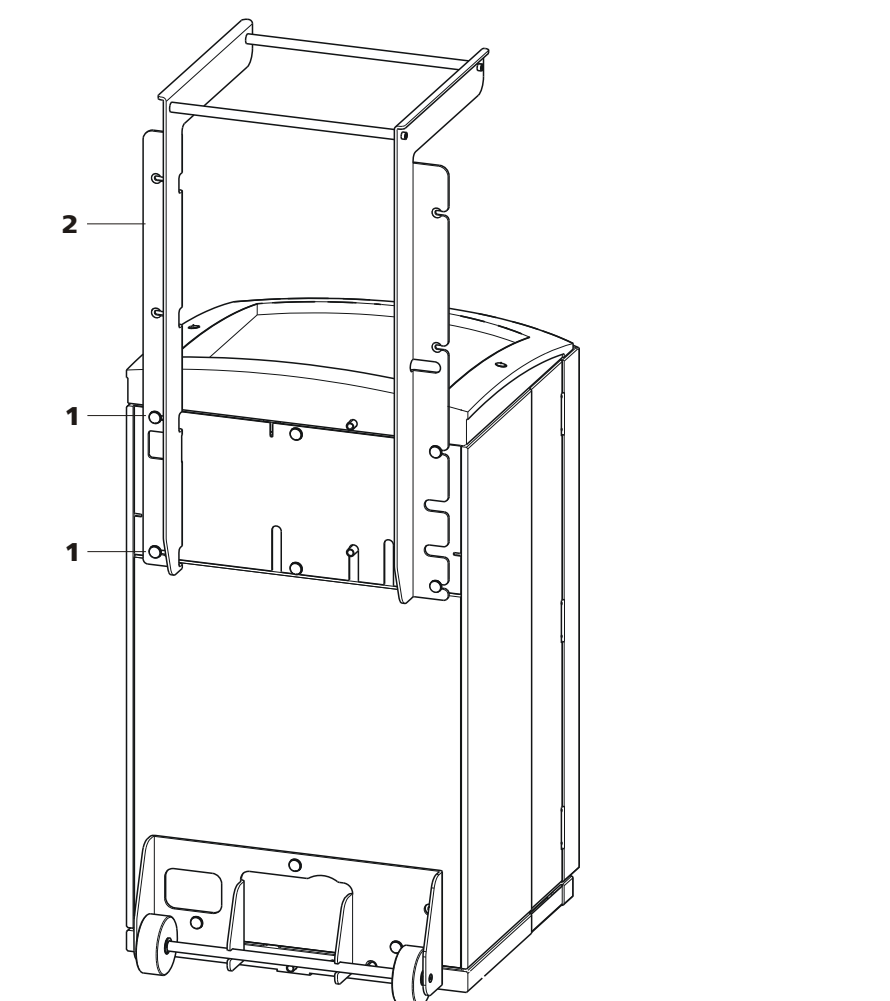


Figure 6 Handle as MPak holder

1 Knurled screws
For fastening the handle (6-2) and the rear panel of the detector chamber.

For fastening the handle (6-**2**) and the rear panel of the detector chamber.

2 Handle
Extended. As holder for MPaks (eluent bag).

Extended. As holder for MPaks (eluent bag).

3.6.2 Positioning and connecting the detector

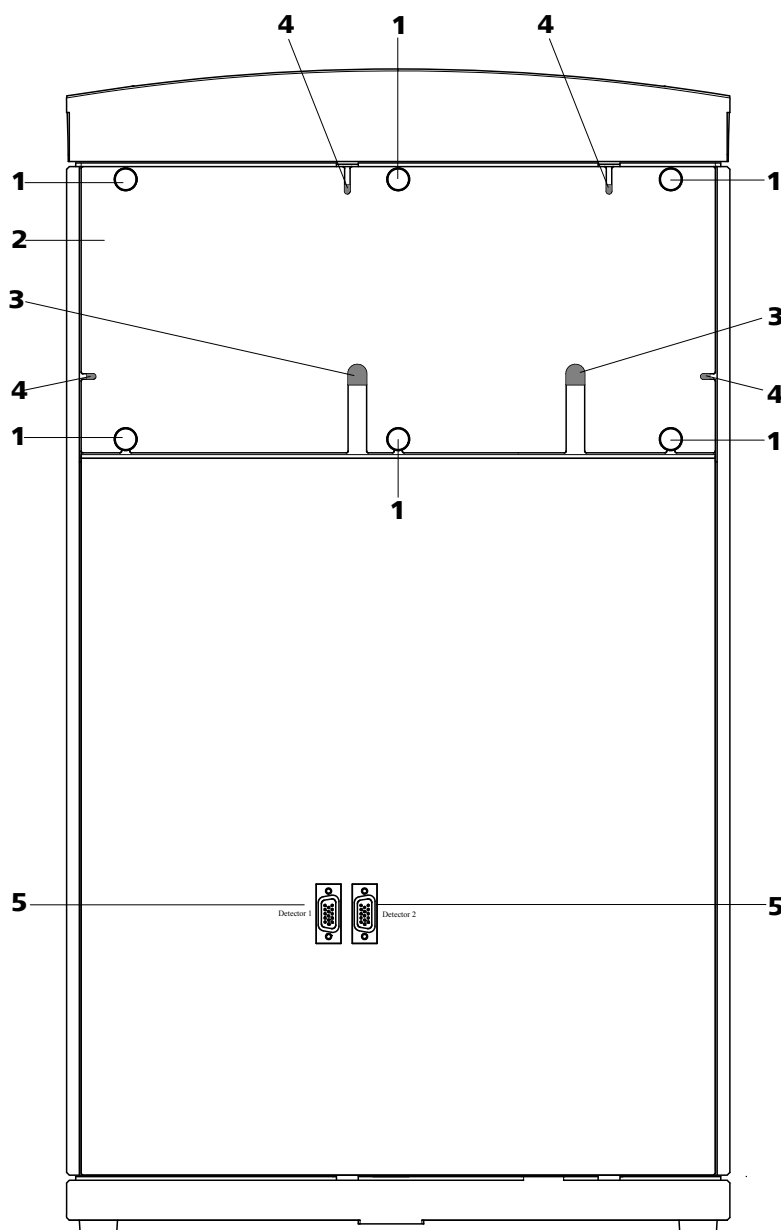


Figure 7 Removable rear panel

- 1 Knurled screws**
For fastening the removable rear panel.

- ### 3 Cable feed-throughs
- For feeding through detector cables.

- 5 Detector connection sockets**
Labeled with *Detector 1* and *Detector 2* for connecting Metrohm detectors.

- 2 Rear panel**
Removable

- #### 4 Capillary feed-throughs



Note

Up to two detectors can be positioned and connected.



Caution

The instrument **must** be turned off when connecting a detector.

1 Removing rear panel

- Unscrew knurled screws (7-1) on the rear panel.
- Remove handle if still fastened to the instrument.
- Remove rear panel (7-2).

2 Positioning the detector

- Put the detector through this opening and position it on the support surface intended for this purpose. Slide it right up to the front.

3 Replacing rear panel

- Insert the detector cable in a cable feed-through (7-3) on the rear panel (7-2).
- Place the detector output capillary in a suitable capillary feed-through.
- Replace (7-2) rear panel.
(Optionally, the handle can be remounted higher up and used as a holder for MPaks.)
- Tighten (7-1) knurled screws.

4 Connecting the detector



Note

The instrument has two detector connection sockets (**7-5**), *Detector 1* and *Detector 2*. You must ensure that the selected connector corresponds to the connector entered in the MagIC Net method.

Recommendation: Use *Detector 1* as standard. In the AnCat system with 2 detectors: *Detector 1* for anions, *Detector 2* for cations.



- Connect the detector cable to the detector connection socket *Detector 1 (7-5)*.

5 Connecting detector output



Note

The detector output capillary must be free of blockages (the measuring cell is tested to 5 MPa = 50 bar backpressure).

Guide the detector output capillary into a sufficiently large waste container and fasten it there.

3.6.3 Transport locking screws

To avoid damage to the high pressure pump and vacuum pump during transport, the pumps are secured with transport locking screws (2-17) (2-15)(2-20).

Remove these transport locking screws before the initial start-up.

Removing transport locking screws

- 1** Remove and keep transport locking screws.



Warning

In order to avoid damage to the pumps, the transport locking screws must be remounted each time the instrument undergoes major transport.

3.6.4 Leak sensor

The leak sensor detects escaping liquid which collects in the base tray of the instrument.

To activate the leak sensor, the leak sensor connector plug (8-2) must be connected, the instrument switched on and the leak sensor switched to **active** in the software.

Connecting the leak sensor

- 1 Plug the leak sensor connector plug (8-2) into the leak sensor connector socket (8-1) on the rear of the instrument (see *Figure 8*, page 27).



Figure 8 Connection for the leak sensor on the rear of the instrument

- ### 3 Leak sensor connection cable
- Is firmly mounted on the rear of the instrument.

3.6.5 Drainage tubing

Fluid that escapes in the covering plate or in the detector chamber flows through the drainage tubing into the base tray and past the leak sensor into the waste container. This ensures that any leaks in the system will be detected by the leak sensor.

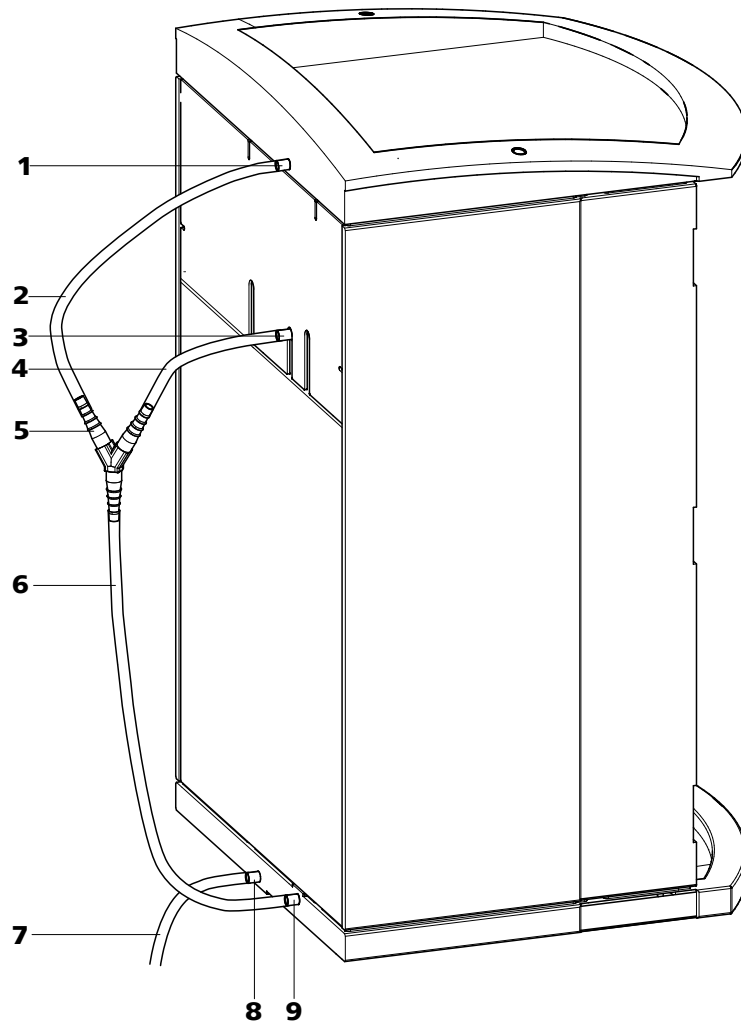


Figure 9 Drainage tubing

- | | |
|---|---|
| <p>1 Drainage tubing connection
For draining escaped fluid from the covering plate.</p> | <p>2 Drainage tubing
Section of the 6.1816.020 silicon tubing. For draining escaped fluid from the covering plate.</p> |
| <p>3 Drainage tubing connection
For draining escaped fluid from the detector chamber.</p> | <p>4 Drainage tubing
Section of the 6.1816.020 silicon tubing. For draining escaped fluid from the detector chamber.</p> |
| <p>5 Y connector 6.1807.010
For connecting the two drainage tubings (9-2) and (9-4).</p> | <p>6 Drainage tubing
Section of the 6.1816.020 silicon tubing. Guides escaped fluid to the leak sensor.</p> |

Section of the 6.1816.020 silicon tubing.
Guides escaped fluid into a waste container.

For draining escaped fluid from the base tray through the connected drainage tubing.

For supplying escaped fluid through the connected drainage tubing to the leak sensor.

Proceed as follows to install the drainage tubing:

Installing drainage tubing

- 1 Plug drainage tubing (9-2) into the drainage tubing connection (9-1) of the covering plate and shorten to required length.
- 2 Plug drainage tubing (9-4) into the drainage tubing connection (9-3) of the detector chamber and shorten to required length.
- 3 Connect drainage tubing (9-2) from the covering plate and drainage tubing (9-4) from the detector chamber to the Y connector (9-5).
- 4 Connect drainage tubing (9-6) to the Y connector (9-5), shorten to required length and plug the other end of the drainage tubing into the drainage tubing connection (9-9).
- 5 Plug drainage tubing (9-7) into the drainage tubing connection (9-8) of the base tray and guide the other end into a waste container.

3.7 Capillary and cable feed-throughs

Several openings have been integrated for feeding through capillaries and cables. They are located on the doors (*see Figure 10, page 30*), on the rear panel (*see Figure 7, Page 24*) or below the covering plate or above the base tray (*see Figure 11, Page 31*).

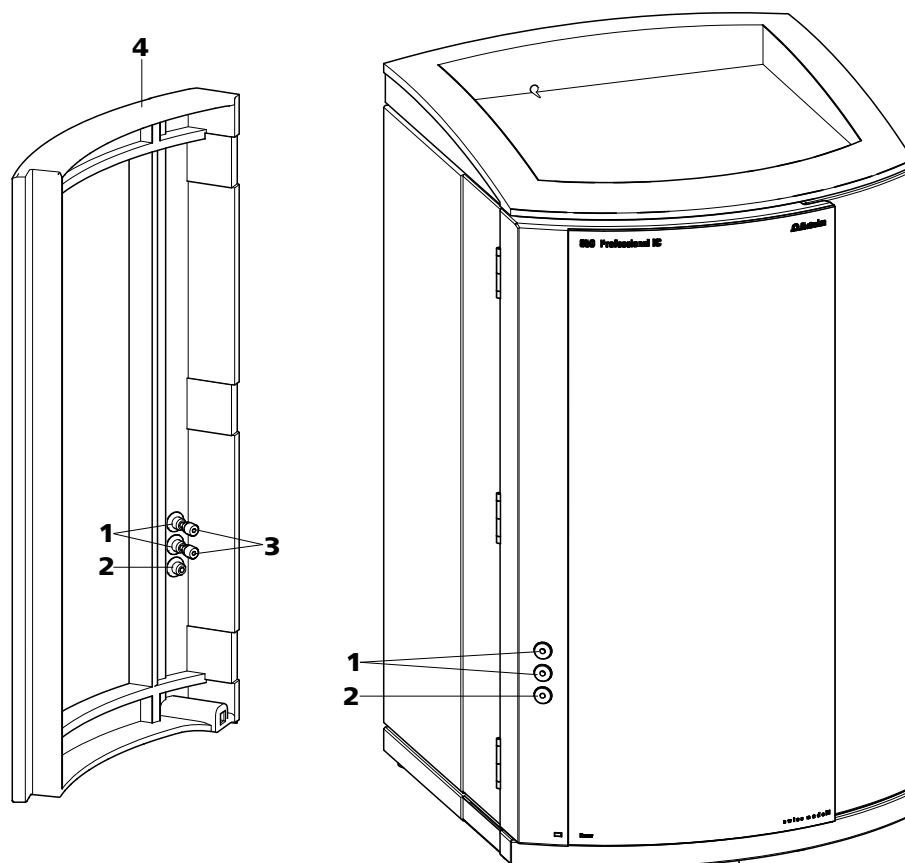


Figure 10 Capillary feed-throughs on the doors

- | | |
|---|---------------------------------|
| 1 Luer connectors
For connecting a 6.2816.020 syringe. For manual sample feeding. | 2 Capillary feed-through |
| 3 PEEK pressure screws, short 6.2744.070 | 4 Door |

Do not feed capillaries through the Luer connectors (10-**1**). The capillaries are fastened with PEEK pressure screws (10-**3**) from inside to the Luer connector. From outside, liquid can be aspirated or injected with a syringe.



1	Side panel (right) Right panel.	2	Rear of the instrument
3	Side panel (left) Left panel.	4	Front of the instrument
5	Capillary feed-through Upper. From front to right.	6	Capillary feed-through Upper. From front to back.
7	Capillary feed-through Upper. From front to back.	8	Capillary feed-through Upper. From front to left.
9	Capillary feed-through Lower. From front to right.	10	Capillary feed-through Lower. From front to back.
11	Capillary feed-through Lower. From front to back.	12	Capillary feed-through Lower. From front to left.

3.8 Eluent

3.8.1 Connecting eluent bottle

The eluent is aspirated out of the eluent bottle via the eluent aspiration tubing (12-**1**).

The eluent aspiration tubing is connected to the eluent degasser (*see Chapter 3.9, page 36*). The tubing must be threaded through a suitable capillary feed-through (*see Chapter 3.7, page 29*) of the instrument before the other end can be equipped.

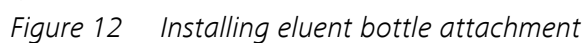
You will require the parts from the following accessories for equipping the eluent aspiration tubing:

- 6.1602.160 eluent bottle attachment GL 45
- 6.2744.210 tubing adapter for aspiration filter
- 6.2821.090 aspiration filter

To equip the eluent aspiration tubing proceed as follows:

Assembling eluent aspiration tubing

- 1 Guide the free end of the eluent aspiration tubing (12-1) out of the instrument through a suitable capillary feed-through.
- 2 **Installing the 6.1602.160 eluent bottle attachment**
 - Slide tubing nipple (12-2) and O-ring (12-3) onto the eluent aspiration tubing (12-1).
 - Push eluent aspiration tubing (12-1) through the bottle attachment (12-4) and screw tight.



- ### 3 Mounting aspiration filter

-

Figure 13 Mounting aspiration filter

- 850 Professional IC

4 Installing tubing weighting and aspiration filter

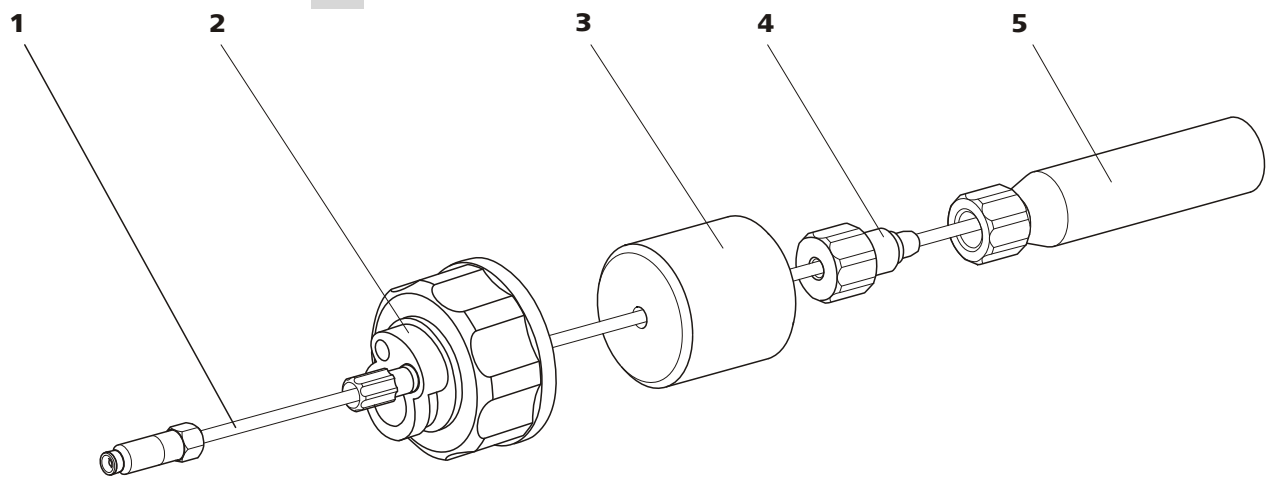


Figure 14 Installing tubing weighting and aspiration filter

1	Eluent aspiration tubing 6.1834.080	2	Eluent bottle attachment 6.1602.160
3	Tubing weighting From accessory set 6.2744.210.	4	Clamping screw From accessory set 6.2744.210.
5	Aspiration filter 6.2821.090 With filter holder from accessory set 6.2744.210.		

- Slide the tubing weighting (14-**3**) onto the eluent aspiration tubing (14-**1**).
- Slide the clamping screw (14-**4**) onto the eluent aspiration tubing (14-**1**).
- Insert eluent aspiration tubing (14-**1**) into the aspiration filter (14-**5**). The end of the tubing must touch the base of the filter.
- Screw together clamping screw (14-**4**) and filter holder (13-**1**). The end of the tubing must still touch the base of the filter.

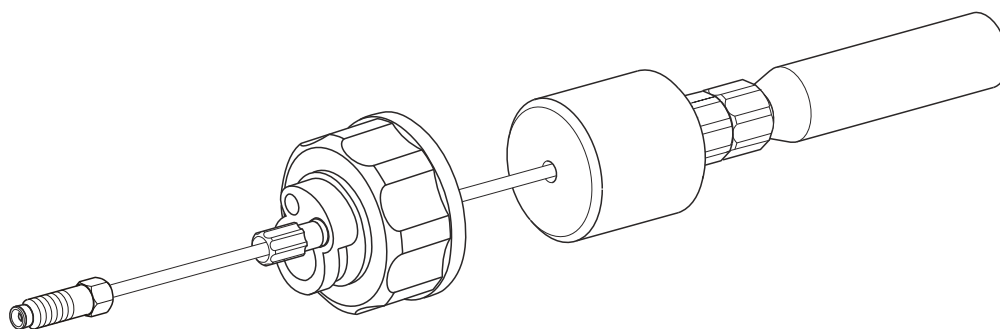


Figure 15 Eluent aspiration tubing fully equipped.

5 Mounting eluent aspiration tubing to the eluent bottle

- Insert the eluent aspiration tubing into the eluent bottle (**16-10**).

- ## 6 Mounting the adsorber tube



- First, place a piece of wadding (16-3), then the CO₂ adsorber (16-4) in the large opening of the adsorber tube (16-2) and close with the plastic cover.
- Fasten the adsorber tube (16-2) using the SGJ clip (16-12) onto the bottle attachment (16-11).

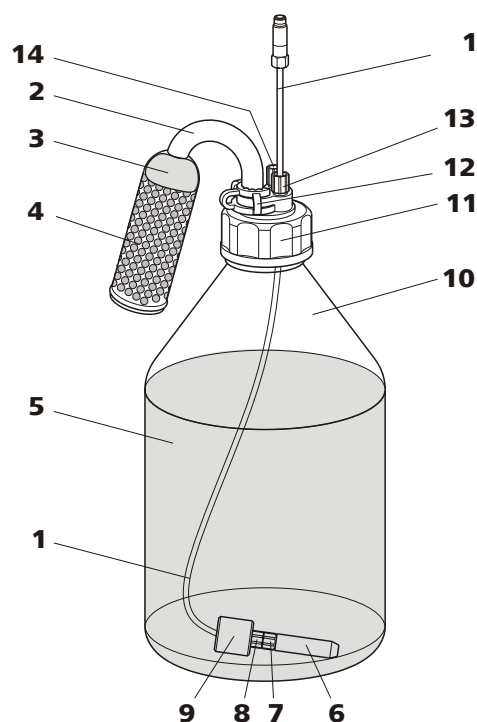


Figure 16 Eluent bottle – connected

1 Eluent aspiration tubing 6.1834.080 For aspirating the eluent. Pre-installed.	2 Adsorber tube 6.1609.000
3 Wadding	4 CO₂ adsorber Adsorbs CO₂ from the air (e.g. Merck soda lime with indicator, no. 6839.10).



Connecting inlet to the high pressure pump



5 Backup ring

Caution

- Plug the eluent aspiration tubing (19-4) into the coupling (19-2).
- Tighten clamping screw (19-3).



3.10.2 Deaerating the high pressure pump

The high pressure pump will only operate perfectly if the pump head contains no more air bubbles. Therefore it must be deaerated during initial start-up and after every change of eluent.



Caution

The high pressure pump must **not** be deaerated before the initial start-up (see *Chapter 4.1, page 71*).

Deaerate the high pressure pump as follows (see Figure 20, page 41):

The instrument must be connected to the PC and switched on to deaerate the high pressure pump.

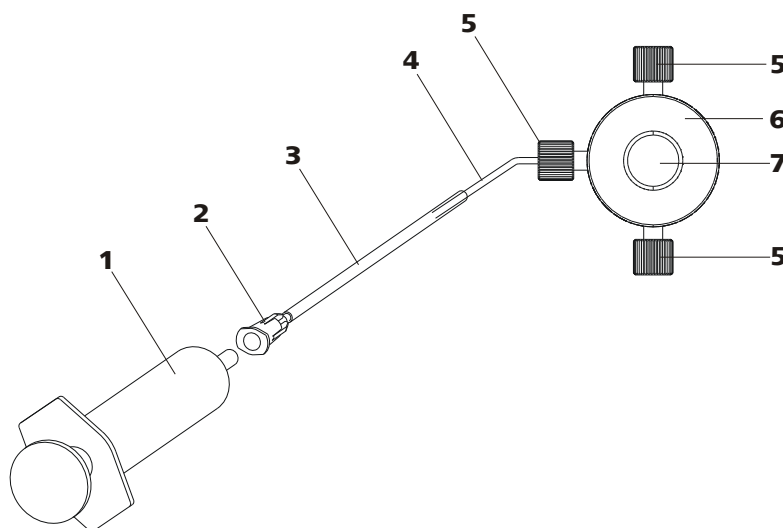


Figure 20 Deaerating the high pressure pump

1	Syringe 10 mL 6.2816.020 For aspirating the eluent.	2	Luer connector On purging needle.
3	Purging needle 6.2816.040	4	Deaerating capillary
5	PEEK pressure screws, short 6.2744.070	6	Purge valve
7	Purge valve rotary knob		

1 Connecting the purging needle

- Push the end of the purging needle (20-**3**) over the end of the deaerating capillary (20-**4**) on the purge valve.

2 Connecting the syringe

- Insert syringe (20-**1**) in the Luer connector (20-**2**) of the purging needle (see Figure 20, page 41).

3 Opening purge valve

- Open the rotary knob (20-7) by approx. ½ rotation counterclockwise.

4 Setting the flow rate

- Start MagIC Net (if not yet started).
- Ensure that the eluent aspiration tubing is immersed sufficiently in the eluent.
- Let the high pressure pump run.

5 Aspirating eluent

- Aspirate with the syringe (20-1) until bubble-free eluent flows into the syringe.

6 Completing deaerating

- Turn off high pressure pump.
- Close rotary knob (20-**7**).
- Remove syringe (20-**1**) from the Luer connector (20-**2**).
- Pull the purging needle (20-**3**) out of the deaerating capillary (20-**4**).

3.11 Inline filter

Between the purge valve and the pulsation damper the 6.2821.120 inline filter is installed as protection against particles.

Inline filters protect the separation column securely against possible contamination from the eluent. Inline filters can however also just as well be used for the purpose of protecting the suppressor against contaminations in the regeneration or rinsing solutions. The fine 2 µm material of the readily and easily replaceable filter platelets removes particles such as bacteria and algae from the solutions.



Note

The inline filter is already installed in the newly delivered instrument. The following installation instructions need **not** be carried out at the time of initial installation.

Installing the inline filter



 **Caution**

Observe the flow direction marked on the filter housing for the connection of the inline filter.



- 4 Connection capillary**
Connects the inline filter with the pulsation damper.

- 1 Screw on the connection capillary running from the purge valve with a 6.2744.070 pressure screw to the input side of the inline filters.
- 2 Screw on the connection capillary running to the pulsation damper with a 6.2744.070 pressure screw to the output side of the inline filter.

3.12 Pulsation damper



 **Note**

The pulsation damper is already installed in the newly delivered instrument.



 **Caution**

The pulsation damper is maintenance-free and may not be opened.

The pulsation damper protects the separation column from damage caused by pressure fluctuations when switching the injection valve, and reduces interfering pulsations during highly sensitive measurements. In order to ensure these functionalities, it must be connected between the high pressure pump (see *Chapter 3.10, page 37*) and injection valve (see *Chapter 3.14, page 46*).

The pulsation damper can be operated in both directions.

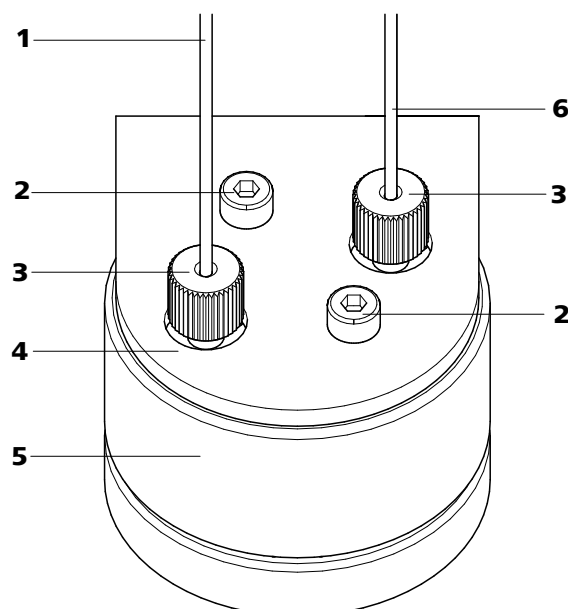


Figure 22 Pulsation damper – Connection

1	Connection capillary Connection to the inline filter.	2	Fastening screws
3	PEEK pressure screws, short 6.2744.070	4	Holder for pulsation damper
5	Pulsation damper 6.2620.150	6	Connection capillary Connection to the injection valve.

3.13 Sample degasser

The sample degasser removes gas bubbles and dissolved gases from the sample. For degassing, the sample flows into a vacuum chamber through a special fluoropolymer capillary.

Gas bubbles in the sample lead to poor reproducibility, as the quantity of sample in the sample loop would not always be the same. Samples (containing gas) should therefore be degassed before injection. For this the sample is sucked through a degasser chamber before injection, whereby any gas bubbles are automatically removed.

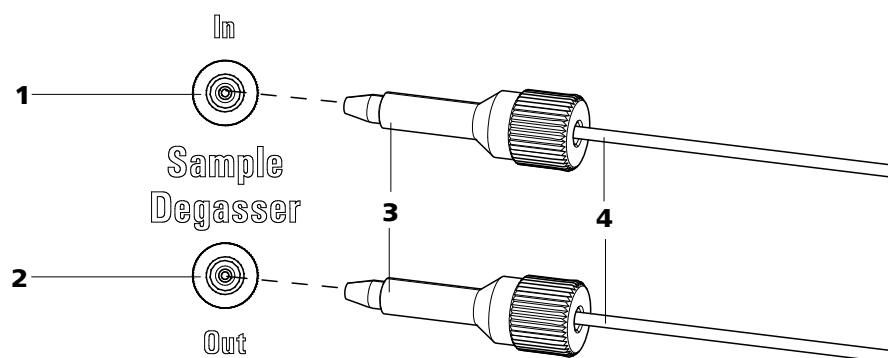
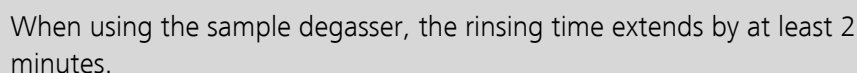


Figure 23 Sample degasser

1	Sample degasser input	2	Sample degasser output
3	PEEK pressure screw, long 6.2744.090	4	Connection capillaries 6.1803.040

Connecting the sample degasser

- 1 Remove and keep 6.2744.220 thread stoppers from the input and output of the sample degasser.
- 2 Connect the end of the 6.1803.040 sample aspirating capillary connected to the injection valve with a long PEEK pressure screw (23-**3**) to the output of the sample degasser (23-**2**).
- 3 Connect 6.1803.040 connection capillary with a long PEEK pressure screw (23-**3**) to the input of the sample degasser (23-**1**).

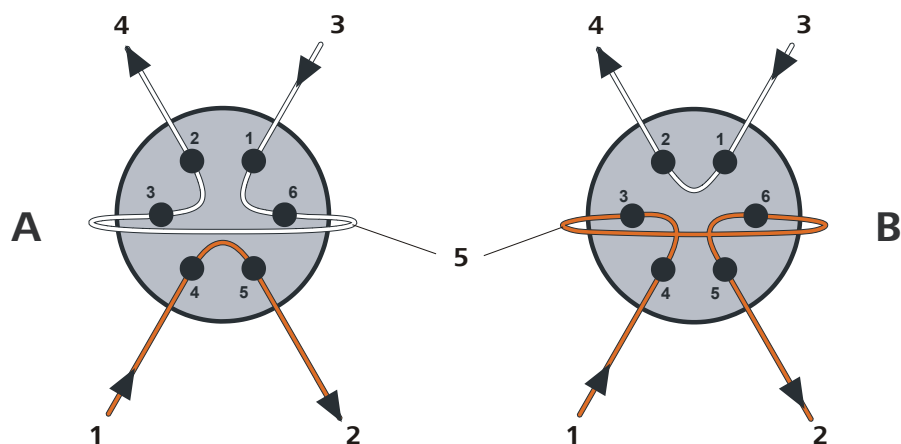


Figure 25 Injection valve – Positions

A	Position FILL	B	Position INJECT
1	Eluent input Capillary coming from the high pressure pump.	2	Eluent output Capillary to the column.
3	Sample input Sample aspirating capillary.	4	Sample output Capillary to waste container.
5	Sample loop		

Position A

In the position **FILL**, the sample solution flows through the sample loop to the waste container. The eluent flows directly to the separation column at the same time.

Position B

In the position **INJECT**, the eluent flows through the sample loop to the separation column. If sample solution is to be found in the sample loop at the time of the valve switchover, then this will be conveyed along with the eluent, thus making its way to the separation column. The flow in the sample path is either stopped or the sample flows directly to the waste container.

3.14.3 Selecting the sample loop

The amount of sample solution injected depends on the volume of the sample loop. The choice is made on the basis of the application. The following sample loops are normally used:

Cation determination	10 µL
Anion determination with suppression	20 µL
Anion determination without suppression	100 µL

3.15 Column thermostat

The column thermostat controls the temperature of the column and eluent channel and thus ensures stable measuring conditions. It provides space for 2 separation columns.

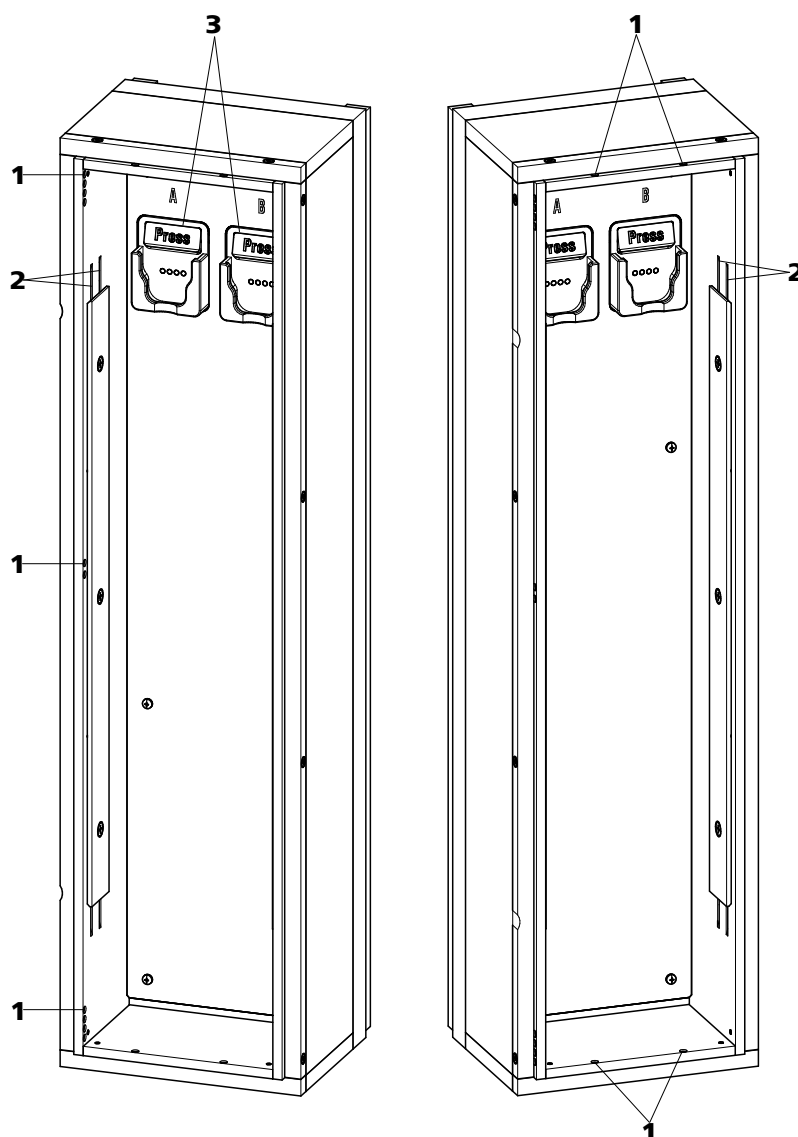


Figure 26 Column thermostat

1 Capillary feed-throughs

For guiding the capillaries in and out.

2 Capillary recesses

For controlling the temperature of the eluent.

Preheating capillary already pre-installed.

3 Column holder

For fastening the columns.

With column recognition.

The column thermostat contains two column holders (26-**3**) equipped with chip recognition. The separation columns can be clicked into the column holder with the chip.

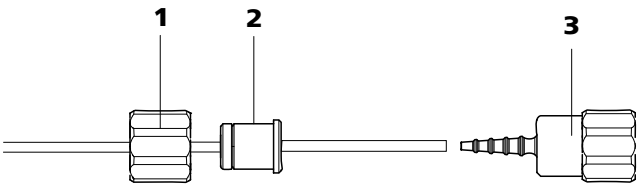


Figure 31 Install pump tubing connection without filter

1	Union nut	2	Adapter
3	Tubing olive		

- Slide union nut (31-1) onto the pump tubing.
- Select a suitable adapter (31-2) and slide it onto the pump tubing. The type of adapter depends on the pump tubing (see Table 1, page 58).
- Place the tubing olive (31-3) onto the pump tubing.
- Screw the union nut (31-1) onto the tubing olive (31-3).

4 Inserting the pump tubing

- Press the contact pressure lever all the way down.
- Place the pump tubing in the tubing cartridge. The stoppers (29-3) must snap into the corresponding holders of the tubing cartridge.

5 Inserting the tubing cartridge

- Hang the tubing cartridge in the mounting pin and press in the cartridge holder until the snap-action lever snaps in.

6 Connecting the capillaries

- Screw the respective capillaries tightly to the two tubing olives with PEEK pressure screws (29-1).

Table 1 Pump tubings and suitable adapters

Pump tubing	Adapter
6.1826.020 (blue/blue)	
6.1826.310 (orange/green)	
6.1826.320 (orange/yellow)	
6.1826.330 (orange/white)	

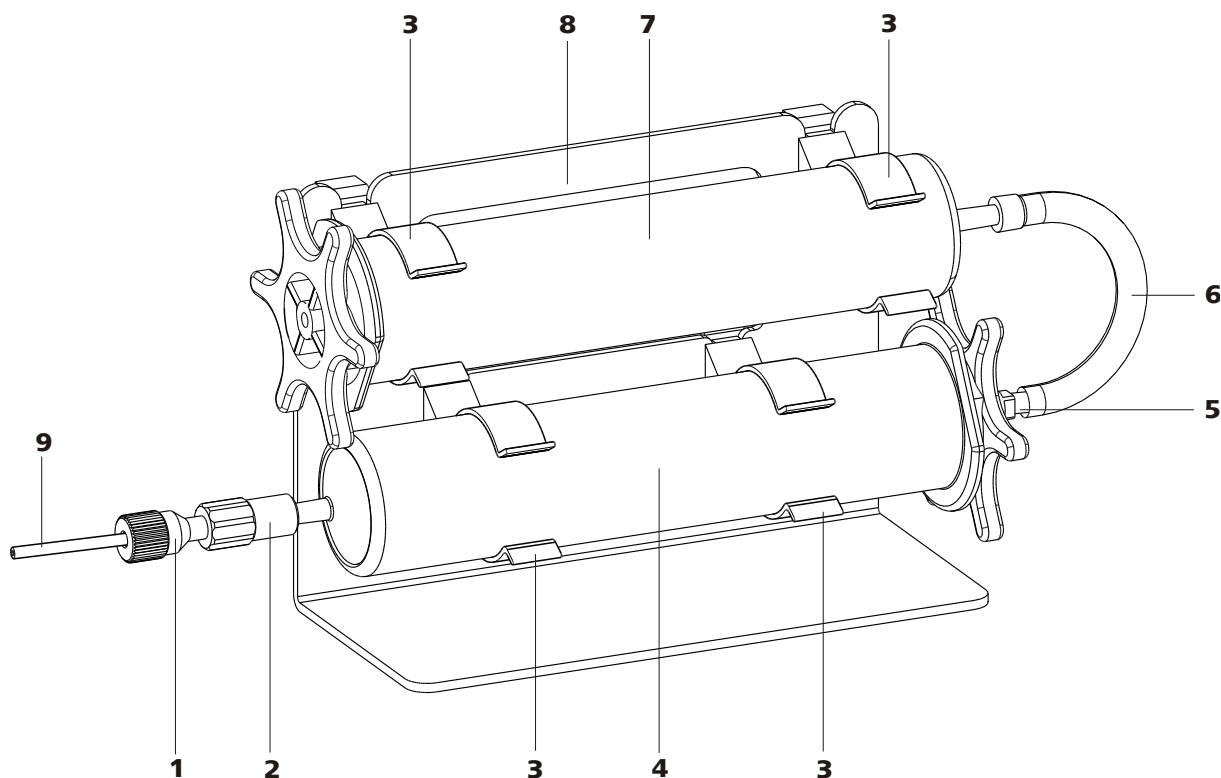


Figure 33 Adsorption cartridge holder

1 PEEK pressure screw, short 6.2744.070

3 Clips
For fastening the adsorption cartridges.

5 Adapter 6.1808.190
For connecting the H₂O adsorption cartridge and CO₂ adsorption cartridge.

7 H₂O adsorption cartridge 6.2837.010
For removing the H₂O from the aspirated air.
Filled with bead desiccant.

9 MCS aspirating capillary
Connection to the MCS. Corresponds to (32-**3**).

2 Luer coupling 6.2744.120

4 CO₂ adsorption cartridge 6.2837.000
For removing the CO₂ from the aspirated air.
3-layer filled, blue-brown-gray.

6 PVC tubing
For connecting the H₂O adsorption cartridge and CO₂ adsorption cartridge.

8 Adsorption cartridge holder 6.2057.080

Installing the adsorption cartridges

1 Preparing the adsorption cartridge holder

Push the 4 clips (33-**3**) into the slot of the adsorption cartridge holder (33-**8**).

2 Removing the caps

- Remove the two locking caps at the tip of the two cartridges.
- In the case of the H₂O adsorption cartridge, replace the round sealing cap on the larger end with the star-shaped sealing cap.

3 Connecting the CO₂ adsorption cartridge

- Insert the CO₂ adsorption cartridge into the coupling (33-2) on the end of the MCS aspirating capillary .
- Click the CO₂ adsorption cartridge into the two lower clips (33-3) of the adsorption cartridge holder (33-8).

4 Connecting the PVC tubing

- Insert the adapter (33-5) into the CO₂ adsorption cartridge.
- Fasten the PVC tubing (33-6) on the adapter (33-5).

5 Connecting the H₂O adsorption cartridge

- Place the H₂O adsorption cartridge into the PVC tubing (33-6).
- Click the H₂O adsorption cartridge into the two upper clips (33-3) of the adsorption cartridge holder (33-8).

6 Placing the adsorption cartridge holder in the instrument

- Place the adsorption cartridge holder with cartridges into the detector chamber of the instrument.

3.19 Conductivity detector

The conductivity detector continuously measures the conductivity of the liquid passing through and indicates these signals in digital form (DSP – Digital Signal Processing). The conductivity detector exhibits outstanding thermal stability and thus guarantees reproducible measuring conditions.

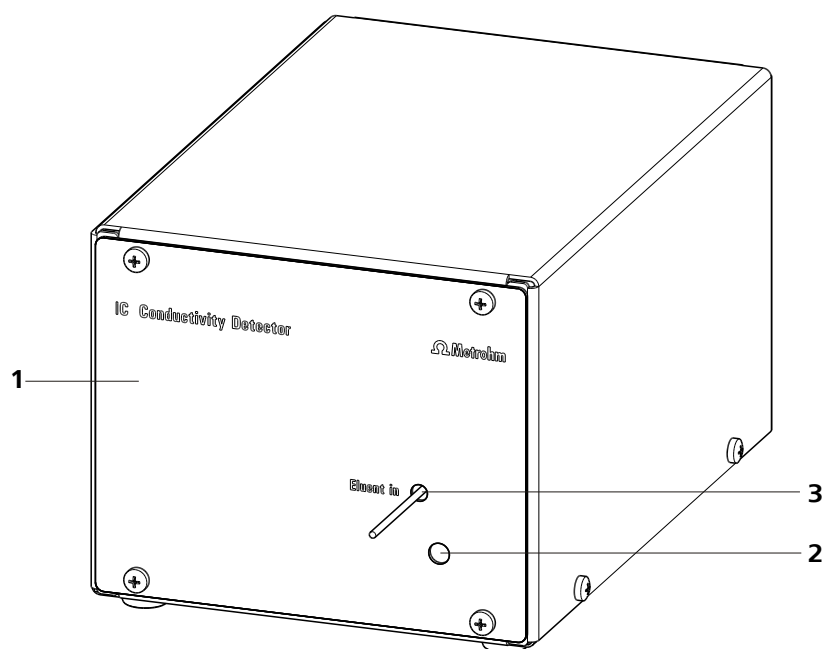


Figure 34 Conductivity detector front

- 1 IC detector 1.850.9010**

- ### 3 Detector input capillary
- Permanently installed.

- ## 2 Opening for temperature sensor



- | | |
|---|--|
| 1 Detector cable
With mounted plug. | 2 Detector output capillary
Permanently installed. |
| 3 Type plate
With serial number. | |



In order to prevent unnecessary peak widening after separation, the connection between the outlet of the separation column and the inlet in the detector should be kept as short as possible.

Connecting the detector input capillary to the MCS

- 1 ■ Fasten the detector input capillary (36-1) using a 2.2744.090 long PEEK pressure screw (36-2) on the output of the MCS (36-3).

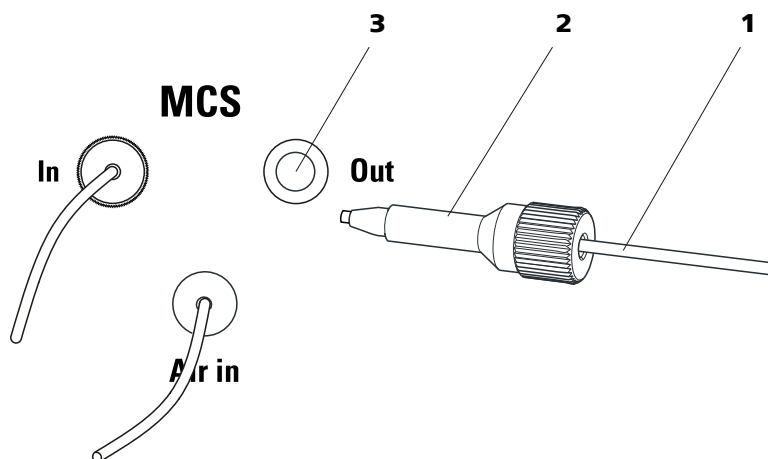


Figure 36 Connection detector – MCS

1 Detector input capillary

2 Pressure screw, long 6.2744.090

3 MCS output

3.20 Connecting the instrument

3.20.1 Connecting the instrument to the PC



Note

The instrument must be turned off when connecting the PC.

1 Connecting the USB cable

Connect the PC connection socket (2-**18**) of the instrument to a USB connector of the computer via the 6.2151.020 USB cable.

3.20.2 Connecting the instrument to mains supply



Warning

The power supply unit must not get wet. Protect it against the direct effect of liquids.

Mains cable

Which mains cable is supplied depends on the location:

- 6.2122.020 with plug SEV 12 (Switzerland, ...)
- 6.2122.040 with plug CEE(7), VII (Germany, ...)
- 6.2122.070 with plug NEMA 5-15 (USA, ...)

1 Connecting the mains cable

- Plug the mains cable into the mains connection socket (2-**4**).
- Connect the mains cable to the mains supply.

2 Switching the instrument on and off

Switch the instrument on and off with the mains switch (2-**3**).

After switching on, the LED on the front of the instrument flashes while a system test is carried out and the connection to the software is established. Once the system test is complete and the connection to the software has been established, the LED lights up continuously.

column

Note

Information regarding which guard column is suitable for your separation column can be found in the **Metrohm IC Column Program** (which is available from your Metrohm agent), the information sheet provided along with your separation column, the product information on the separation column at <http://www.metrohm.com> (product area Ion Chromatography), or obtained directly from your agent.

 **Caution**

New guard columns are filled with solution and are sealed on both sides with stoppers or caps, respectively. Before using the guard column, you need to ensure that this solution is miscible with the eluents used (observe manufacturer's data).



Note

The guard column may only be installed after the **initial start-up** (see Chapter 4.1, page 71) of the instrument. Until then, use the 6.2744.040 coupling instead of the guard and separation column.

Connecting and rinsing the guard column

1 Connecting the guard column



Caution

When inserting the guard column, always ensure that it is inserted correctly corresponding to the flow direction (if indicated).

- Remove locking caps and/or stoppers from the guard column.
- Fasten the lower end of the guard column with a 6.2744.070 short PEEK pressure screw to the column input capillary .
- Fasten the connection capillary (3.4224.240) enclosed with the guard column to the upper end of the guard column with a 6.2744.070 short PEEK pressure screw.

Guard columns are also available which can be screwed with the upper end directly onto the separation column.

2 Rinsing the guard column

- Place beaker under the outlet capillary of the guard column.
- Start the high pressure pump and rinse the guard column approx. 5 minutes with eluent. Set the flow according to the corresponding column information sheet.
- Switch off the high pressure pump again.

3.22 Separation column

The intelligent separation column (iColumn) is the heart of the ion chromatographic analysis. It separates the different components corresponding to their interactions with the column. Metrohm analytical columns are equipped with a chip on which their technical specifications and their history (first use / setting up, operating hours, injections, ...) are saved.



Note

Information regarding which separation column is suitable for your application can be found in the **Metrohm IC Column Program**, the product information for your separation column at <http://www.metrohm.com> in the product area ion chromatography, or obtained directly from your agent.



Caution

New separation columns are filled with solution and are sealed on both sides with stoppers. Before using the column, you need to ensure that this solution is miscible with the eluents used (observe manufacturer's data).

You can find the separation columns and guard columns currently available from Metrohm in the Metrohm IC Column Program, or in the Internet at <http://www.metrohm.com> in the product area Ion Chromatography. A test chromatogram and an information sheet are provided along with each column. You can request detailed information on special IC applications in the corresponding "**Application Bulletins**" or "**Application Notes**", available in the Internet at <http://www.metrohm.com> in the Applications area or via the Metrohm agent responsible free of charge.



Note

The separation column may only be installed after the **initial start-up** (see Chapter 4.1, page 71) of the instrument. Until then, use the 6.2744.040 coupling instead of the guard and separation column.

Connecting and rinsing the separation column

1 Connecting the separation column



Caution

When using the columns, always ensure that these are correctly inserted corresponding to the flow direction indicated.

- Remove stoppers from the separation column.
- Connect the lower end of the separation column with a 6.2744.070 short PEEK pressure screw to the outlet capillary of the guard column (if used), or to the column input capillary.

2 Rinsing the separation column

- Place beaker under the outlet end of the separation column.
- Set the flow rate of the high pressure pump to the value suitable for the separation column selected.
- Start the high pressure pump and rinse the separation column approx. 10 minutes with eluent.
- Switch off the high pressure pump again.

3 Mounting the separation column

- Hang separation column with chip into the column holder.



Note

The iColumns are equipped with a chip on which their operating data is saved. The chip has to be hooked into the chip holder provided for this so that the column recognition can function.

4 Start-up

The chapter *Start-up* is divided into 2 sections:

Initial start-up

The **initial start-up** is carried out during the **initial installation**.

Conditioning

Conditioning is carried out as a final installation step and each time after the system is started.

4.1 Initial start-up

The initial start-up is carried out during the initial installation. The entire system is flushed before guard column and separation column are installed.



Caution

The separation column and guard column may not be installed for the initial start-up.

Make sure that the 6.2744.040 coupling is being used instead of the columns.

Perform the following steps during the initial start-up:

1 Preparing the software

- Start the PC program **MagIC Net**.
- Open the **Equilibration** tab in **MagIC Net**.
- Select (or create) a suitable method.

2 Preparing the instrument

- Ensure that the eluent aspiration tubing is immersed in the eluent and that there is enough eluent in the eluent bottle.
- Switch on the instrument.

3 Deaerating the high pressure pump

- Deaerate the high pressure pump(s) via the purge valve (*see Chapter 3.10.2, page 40*).

2 Preparing the instrument

- Ensure that the column is correctly mounted in accordance with the flow direction indicated on the label (arrow must point in the direction of flow).
- Ensure that the eluent aspiration tubing is immersed in the eluent and that there is enough eluent in the eluent bottle.

3 Checking leak-tightness

- In **MagIC Net**, start the equilibration.
- Check all capillaries and their connections from the high pressure pump to the detector block for signs of liquid escaping. If eluent escapes anywhere, tighten the corresponding pressure screw or loosen the connection, check the end of the capillary, shorten it with a capillary cutter if necessary, and restore the connection.

4 Conditioning the system

Rinse the system with eluent until the required stability of the baseline is attained (normally 30 minutes).

The instrument is now ready for measuring samples.

5.1.3 Operation



Caution

In order to avoid disturbing temperature influences, the entire system including the eluent bottle must be protected against direct sunlight.

5.1.4 Shutting down

If the instrument is shut down for a longer period, the entire IC system (without separation column) must be rinsed salt-free with methanol/ultra pure water (1:4), in order to prevent eluent salts from forming crystals which may cause subsequent damage.

Guard column and separation column are excluded from the eluent path for rinsing. The connection capillaries are directly connected to one another with a 6.2744.040 coupling. Rinsing is carried out with methanol/ultra pure water (1:4) until the conductivity drops below 10 $\mu\text{S}/\text{cm}$.

Rinse with eluent for at least 15 minutes at starting up again and before connecting the guard column and separation column.

5.2 Capillary connections

5.2.1 Operation

All connections between injection valve (*see Chapter 3.14, page 46*), separation column (*see Chapter 3.22, page 69*) and detector (*see Chapter 3.19, page 63*) must be as short as possible, have a low dead volume and be completely leak-tight. The PEEK capillary after the detector must be free of blockages (the measuring cell is tested to 5 MPa = 50 bar back-pressure). Only use PEEK capillaries with an internal diameter of 0.25 mm in the high pressure range between the high pressure pump (*see Chapter 3.10, page 37*) and the detector.



5.3 Door



Caution

The door is made of PMMA (polymethylmetacrylate). It must never be cleaned with abrasive media or solvents.



Caution

Never use the door as a handle.

5.4 Eluent

5.4.1 Production

The chemicals used for the production of eluents should have a degree of purity of at least "p.a.". Only ultra pure water (resistance > 18.2 MΩ*cm) may be used for dilution (this generally applies for reagents which are used in ion chromatography).

Newly produced eluents should always be microfiltered (filter 0.45 μm).



Caution

Only microfiltered (filter 0.45 μm) eluents may be used.

The composition of the eluent has a crucial effect on the chromatographic analysis:

Concentration	An increase in the concentration generally leads to shorter retention times and faster separation, but also to higher background conductivity.
pH	pH changes result in shifts in the dissociation equilibria and hence changes in the retention times.
Organic solvents	The addition of an organic solvent (e.g. methanol, acetone, acetonitrile) to aqueous eluents generally accelerates lipophilic ions.

5.4.2.1 Supply bottle

Moreover, condensation must also be prevented in the eluent bottle. Drop formation can change the concentration ratio in the eluent.

To protect the IC system against foreign particles, we recommend aspirating the eluents via a 6.2821.090 aspiration filter (*13-2*). This aspiration filter must be replaced should it show signs of yellow discoloration (but no later than every 3 months).

In the case of very sensitive measurements, the eluent should be stirred constantly with a magnetic stirrer.

5.4.2.3 Changing the eluent

When changing the eluent, it must be ensured that no precipitates can occur. Solutions following one another in direct succession must therefore be miscible. If the system has to be rinsed organically, several solvents with rising or falling lipophilia must be used.

5.5 High pressure pump

5.5.1 Protection



Caution

The pump head is filled ex works with methanol/ultra pure water. It must be ensured that the eluent used is freely miscible with the solvent remaining in the pump head.

To protect the high pressure pump against **foreign particles**, we recommend that the eluent undergoes a **microfiltration** (filter 0.45 µm) before being aspirated via a 6.2821.090 aspiration filter (*see "Assembling eluent aspiration tubing", page 32*).

Salt crystals between the piston and seal cause abrasion particles which can find their way into the eluent. These lead to contaminated valves, a rise in pressure and in extreme cases scratched pistons. It is therefore essential to ensure that **no precipitates** can occur (*see Chapter 5.4.2.3, page 77*).



Caution

In order to spare the pump seals, the pump should not be operated dry. Therefore ensure that the eluent supply is correctly connected and that there is enough eluent in the eluent bottle each time before turning on the pump.

5.5.2 Maintenance



Caution

Maintenance work on the high pressure pump may not be carried out unless the **instrument is switched off**.

Pump head maintenance

An unstable baseline (pulsation, flow fluctuations) is in many cases the result of contaminated valves (43-**2**), (43-**3**) or defective, leaking piston seals on the high pressure pump. Proceed as follows for cleaning contaminated valves and/or replacing worn parts such as pistons, piston seal and valves:

This maintenance work should be carried out at least once a year.

Removing the pump head

- 1 Turn off high pressure pump and wait until pressure is released.
- 2 Loosen the pressure screw on the inlet valve holder (18-**2**) and unscrew the coupling (18-**9**), the pump head input capillary (18-**7**) and the eluent aspiration tubing from the pump head.
- 3 Unscrew the pump head output capillary (18-**13**) from the pump head.
- 4 Remove pump head from the pump housing by loosening the 4 fastening screws (18-**5**) using the 6.2621.030 hexagon key. The main piston is on the left (viewed from the front), and the auxiliary piston is on the right.



- Remove the backup ring from the pump head and lay to the other parts.

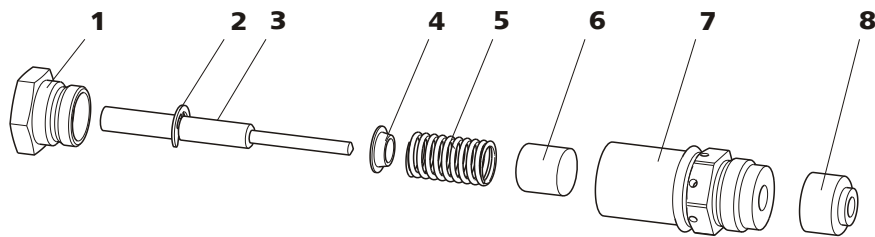


Figure 38 Components of the piston cartridge

1	Piston cartridge screw	2	Retaining washer
3	Zirconium oxide piston with piston shaft Order number: 6.2824.070.	4	Spring retainer
5	Spring Order number: 6.2824.060.	6	Inner plastic sleeve Protects from metallic abrasion.
7	Piston cartridge	8	Backup ring

3 Cleaning the components of the piston

- Clean zirconium oxide pistons contaminated by abrasion or deposits with pure abrasive cleaning powder, rinse particle free with ultra pure water and dry.
Replace highly contaminated or scratched zirconium oxide pistons (spare part: 6.2824.070 zirconium oxide piston).
- Rinse the other parts of the piston and dry with a lint-free cloth.

4 Assembling the piston

- Insert the inner plastic sleeve, spring and spring retainer into the piston cartridge.
- Slide the zirconium oxide piston carefully into the piston cartridge until its tip emerges from the small opening of the piston cartridge.
- Attach screw and tighten by hand.

Replacing the piston seal

The 6.2617.010 special tool (see Figure 39, page 81) is necessary in order to remove the piston seal from the pump head. It consists of two

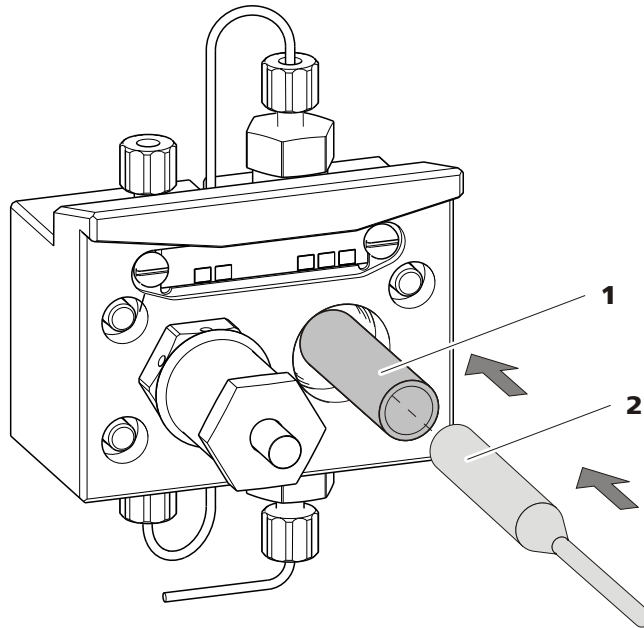


Figure 42 Inserting the piston seal into the pump head

4 Replacing the piston cartridge

Screw the assembled piston cartridge back into the pump head and tighten, first by hand, then additionally by approx. 15° with a wrench.

Cleaning the inlet valve and outlet valve

1 Removing valves

- Unscrew the connection capillary for the auxiliary piston (18-1) from the outlet valve holder.
- Unscrew the holders for the inlet and outlet valves and remove valves.

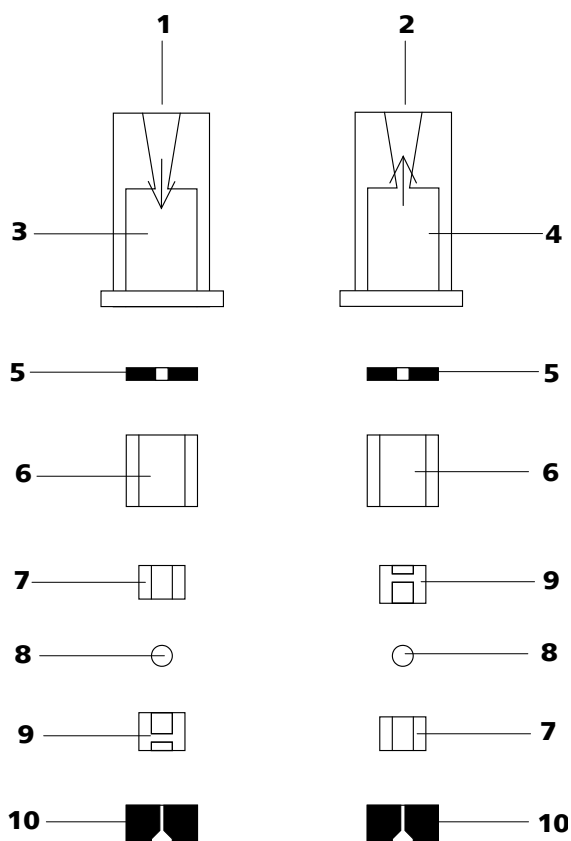


Figure 45 Components of the inlet valve and outlet valve

1	Inlet valve 6.2824.170	2	Outlet valve 6.2824.160
3	Inlet valve housing	4	Outlet valve housing
5	Sealing ring (black)	6	Sleeve
7	Sapphire sleeve The shiny side must point to ruby ball.	8	Ruby ball
9	Ceramic holder for ruby ball	10	Seal The larger opening must point outwards.

4 Clean the components of the valve

Rinse the valve components with ultra pure water and/or acetone and dry with a lint-free cloth.

5 Reassemble the valve

Reassemble valve components *according to figure 45, page 86*.

- Insert the seal with the larger opening faced downwards into the recess of the tool.
- Lay the other valve components above another in the correct sequence (*see Figure 45, page 86*).

- ## 6 Checking the flow direction

If this is not the case, the valve has to be dismantled again and be assembled correctly (see Figure 45, page 86).

Caution

When inserting the valves, please take into account that the liquid is being pumped through the pump head from bottom to top.

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Mounting the pump head



Note

To prevent the pump head from being positioned the wrong way, it is provided with different bore hole depths for the fastening bolts, i. e. a fastening bolt is longer than all others. The bore hole with the greatest depth must therefore be assigned to the longest bolt. If this is not the case, the pump will not function perfectly.

- 1** Mount the pump head on the pump again using the four fastening screws (18-5). Firmly tighten the screws with the 6.2621.030 hexagon key.
- 2** Screw connection capillaries (18-1), (18-7) and (18-13) onto the pump head again.

5.6 Inline filter

5.6.1 Maintenance

The 6.2821.120 inline filters comprise the filter housing (46-**2**), the filter screw (46-**4**) and the filter (46-**3**). New filters (46-**3**) are available under the order number 6.2821.130 (10 items).

The 6.2821.130 filters (21-**3**) should be changed every 3 months (more frequently at higher backpressure).

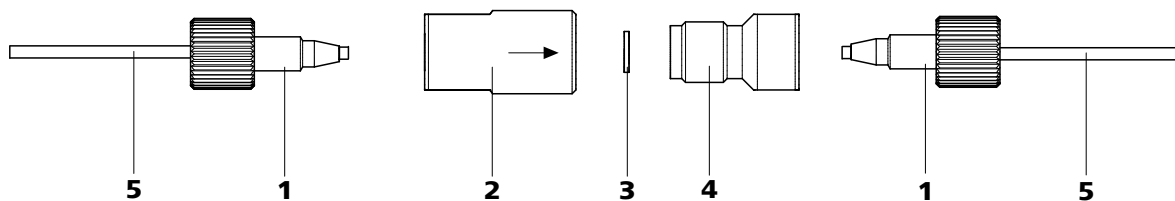


Figure 46 Changing the filter

- 1 PEEK pressure screws, short**
6.2744.070

- 3 6.2821.130 filter**
Packaging contains 10 items.

- ## 5 Connection capillaries

- 2 Filter housing**
Housing of the inline filter. Part of the 6.2821.120 accessories.

- #### 4 Filter screw
- Screw of the inline filter. Part of the 6.2821.120 accessories.

Changing the filter

The flow must be stopped before changing the filter.

1 Removing the inline filter

- Unscrew the pressure screws (46-1) from the inline filter.

2 Unscrewing the filter screw

- Screw the filter screw (46-4) out of the filter housing (46-2) with the aid of two 6.2621.000 adjustable wrenches.

3 Inserting the filter

- Remove the old filter (46-3) with tweezers.
- Place the new filter (46-3) flat in the filter housing with tweezers (46-2).

4 Mounting filter screw

- Screw the filter screw (46-4) back into the filter housing (46-2) and tighten by hand. Then additionally tighten slightly with two 6.2621.000 adjustable wrenches.

5 Remounting the inline filter

- Screw the pressure screws (46-**1**) back onto the inline filter.

6 Rinsing the inline filter

- Dismantle the guard column (if present) and the separation column and replace with a 6.2744.040 coupling.
- Rinse the instrument with eluent.



5.7 Inline sample preparation

To protect the separation column (see Chapter 3.22, Page 69) against foreign particles which can affect the separating efficiency, we recommend that all samples undergo a microfiltration (filter 0.45 μm). The ultrafiltration cell can be used for **filtration** (see documentation on the *IC Equipment for Ultrafiltration*).

Samples containing significant amounts of **gas** should be degassed. The sample degasser (*see Chapter 3.13, Page 45*) (if available) is used for degassing.

Matrix-loaded samples (e.g. blood, oil) should be prepared for the measurement by means of dialysis (see documentation on the *IC Equipment for Dialysis*).

If the concentration of the sample is too high, it should be **diluted** before feeding (see documentation on the *IC Equipment for Sample Dilution*).

For the sample preparation methods **Neutralization** (replacement of e.g. Na^+ with H^+) and **cation exchange** (replacement of e.g. heavy metals with H^+), a sample preparation module (SPM) is used.

5.8 Rinsing the sample path

Before a new sample can be measured, the sample path must be rinsed with it so that the measuring result is not falsified by the previous sample (**Sample carry-over**).

In the case of automated sample feeding, the rinsing time should be at least 3 times the **transfer time**. The transfer time is the time required by the sample to flow from the sample vessel to the end of the sample loop.

Ascertaining the transfer time

The transfer time depends on the pump capacity of the peristaltic pump, the total capillary volume and the volume of the gas removed by the sample degasser (if used) - in other words the amount of gas in the sample.

1 Emptying the sample path

Pump air through the sample path (pump tubing, tubing connections, capillary in the degasser, sample loop) for several minutes until all liquid is displaced by the air.

2

The time measured corresponds to the "transfer time". The rinsing time should be at least 3 times the transfer time.

Checking the rinsing time

It is possible to determine whether the rinsing time is adequate via a direct measurement of the sample carry-over. Proceed as follows:

1

- **Sample A:** A typical sample for the application.
- **Sample B:** Ultra pure water.

2

Let "Sample A" pass through the sample path for the duration of the rinsing time, then inject and measure.

3

Let "Sample B" pass through the sample path for the duration of the rinsing time, then inject and measure.

4

The degree of the sample carry-over corresponds to the ratio of the peak areas of the measurement for sample B to the measurement for sample A. The lower the ratio, the lower the sample carry-over. This ratio can be altered by varying the rinsing time – thus allowing the rinsing time required for the application to be ascertained.

5.9 Sample degasser

5.9.1 Operation

If you are working with sample degassing, the longer "transfer time" (see *Ascertaining the transfer time, page 90*) means that it is also necessary to rinse for longer (with the subsequent sample). The rinsing time should be at least 3 times the "transfer time" so as to minimize the carry-over effects. The "transfer time" itself depends on the pump capacity, total capillary volume and volume of gas removed (i.e. the amount of gas in the sample).



Note

When using the sample degasser, the rinsing time extends by at least 2 minutes.

5.10 Injection valve

5.10.1 Protection

To prevent contamination of the injection valve, a 6.2821.120 inline filter (see Chapter 3.11, page 42) should be mounted between the high pressure pump and the pulsation damper.

5.11 High Capacity Metrohm Suppressor Module (MSM-HC)

5.11.1 Protection

To protect the MSM-HC against foreign particles or bacterial growth, a 6.2744.180 pump tubing connection with filter (*see Figure 30, page 57*) must be mounted between the peristaltic pump and the inlet capillaries of the MSM-HC.

5.11.2 Operation



Note

The suppressor units must never be regenerated in the same flow direction in which the eluent is pumped. Therefore always mount the inlet and outlet capillaries according to diagram outlined in *figure 27*.



Caution



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5.11.3.1 Regenerating the MSM-HC

Regenerating the MSM-HC

- Disconnect the MSM-HC from the separation column and MCS or the detector.

2 Regenerating the MSM-HC



Caution

The pump tubings made of PVC must not be used for rinsing with solutions containing organic solvents. In this case, other pump tubings must be used for rinsing.



Note

The high pressure pump can be used for regeneration. For this, remove the guard column and separation column and connect the capillary directly to the MSM-HC (regenerate in opposite direction).

- Rinse the 3 suppressor units with the following solutions for about 15 minutes each:
 - **Contamination with heavy metals:**
1 mol/L H_2SO_4 + 0.1 mol/L oxalic acid
 - **Contamination with organic cationic complexing agents:**
0.1 mol/L H_2SO_4 / 0.1 mol/L oxalic acid / acetone 5%
 - **Severe contamination with organic substances:**
0.2 mol/L H_2SO_4 / acetone $\geq 20\%$

3 Connecting the MSM-HC to the IC system

- Reconnect the MSM-HC to the IC system. If the capacity problems remain, the MSM-HC rotor A must be replaced (*see Chapter 5.11.3.3, page 97*).

5.11.3.2 Cleaning the MSM-HC

It may be necessary to clean the MSM-HC in the following cases:

- Increased backpressure onto the connection tubings of the MSM-HC.
- Blockage of the MSM-HC which cannot be eliminated (solutions can no longer be pumped through the MSM-HC).
- Jamming of the MSM-HC which cannot be eliminated (MSM-HC can no longer be switched over).

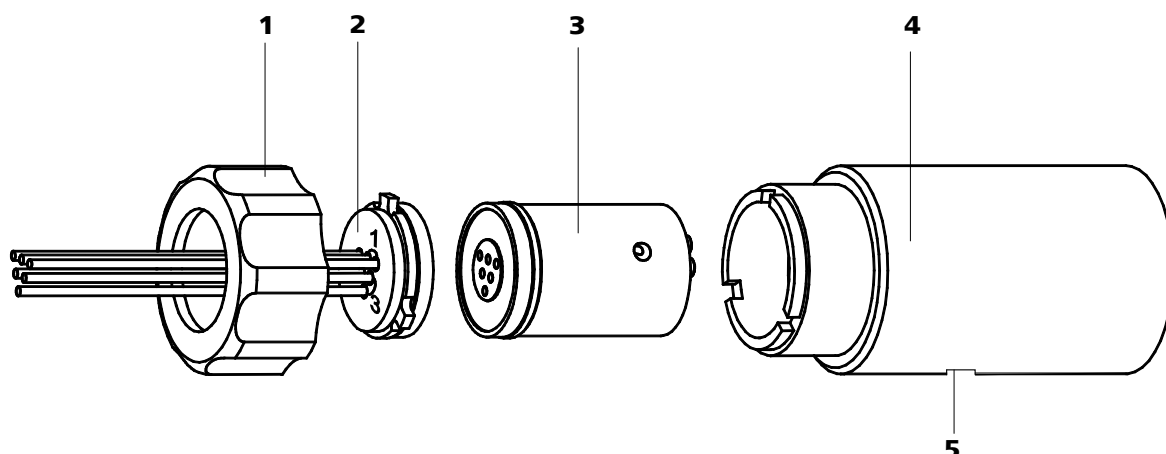


Figure 47 MSM-HC – Components

1	Union nut	2	MSM-HC connecting piece 6.2835.010
3	MSM-HC rotor A 6.2842.000	4	MSM-HC housing
5	Slot in the MSM-HC housing		

Cleaning the MSM-HC

Clean the MSM-HC as follows:

1 Disconnecting the MSM-HC from the IC system

- Switch off the instrument.
- Disconnect the MSM-HC from the separation column, peristaltic pump and detector.

2 Dismantling the MSM-HC

- Unscrew union nut (47-1) from the MSM-HC housing (47-4).
- Pull the MSM-HC connecting piece (47-2) and the MSM-HC rotor A (47-3) out of the MSM-HC housing (47-4). The connecting piece and the MSM-HC rotor A normally stick to one another – if this is not the case: Take a sharp object, insert into the slot (47-5) in the MSM-HC housing, and pull out the MSM-HC rotor A (47-3) in this way.
- Detach the MSM-HC connecting piece (47-2) from MSM-HC rotor A (47-3).

3 Cleaning the supply and discharge lines

- Connect in turn each of the 6 capillary tubings fastened on the MSM-HC connecting piece (47-2) on the high pressure pump (see Chapter 3.10, page 37) and pump through ultra pure water.

3 Mounting filter screw

- Screw the filter screw (48-3) back into the tubing olive (48-1) and tighten by hand. Then additionally tighten with two 6.2621.000 adjustable wrenches.

5.13 Metrohm CO₂ suppressor (MCS)

5.13.1 Replacing the CO₂ adsorption cartridge

The CO₂ adsorption cartridge 6.2837.000 (33-4) must be replaced regularly. This is because of blockages and losses in capacity.

Blockage

Moisture can block the CO₂ adsorption cartridge. This is revealed by a change in color of the cartridge material (the orange part becomes colorless). As the air flow is reduced, the vacuum becomes lower. To protect the CO₂ adsorption cartridge, an H₂O adsorption cartridge (33-7) is installed upstream. Regular regeneration (see Chapter 5.13.2, page 101) of the H₂O adsorption cartridge extends the service life of the CO₂ adsorption cartridge.

Capacity loss

The adsorption capacity of the CO₂ adsorption cartridge is limited. Depending on the period of operation and laboratory environment, the adsorption capacity will diminish over time. This is manifested in a rising baseline (as more CO₂ reaches the detector).

5.13.2 Regenerating the H₂O adsorption cartridge

The function of the H₂O adsorption cartridge is to protect the CO₂ adsorption cartridge against moisture. The service life of the H₂O adsorption cartridge depends on the moisture content of the ambient air. Moisture reduces the capacity of the H₂O adsorption cartridge (which can be observed by a change in color). Before the color changes in the entire filling material (from orange to colorless, with Fluka Order No. 94098), the H₂O adsorption cartridge should be regenerated (see information sheet). The filling material is replaced during regeneration:

- 1 Allow material to dry loose (not in cartridge) at 140 °C overnight and refill. Or dispose of the old material, and fill with new material.
- 2 Cover the packed material with wadding.

5.15.2 Protection

To protect the separation column against foreign particles, which can affect the separating efficiency, we recommend that both the eluent and the samples undergo a microfiltration (filter 0.45 µm) before being aspirated via the 6.2821.090 aspiration filter.

The use of guard columns (*see Chapter 3.21, page 67*) serves for protecting the actual separation columns and increases their service life considerably. Information regarding which guard column is suitable for your separation column can be seen in the **Metrohm IC Column Program** (which is available from your Metrohm agent), the information sheet provided along with your separation column, the product information on the separation column at <http://www.metrohm.com> (product area Ion Chromatography) or can be obtained directly from your agent.

The pulsation damper (*see Chapter 3.12, page 43*) must be installed in order to protect the column material from pressure concussion caused by injection.

5.15.3 Storage

Always store the separation columns sealed and filled according to the data of the column manufacturer when not using them.

5.15.4 Regeneration



Note

The regeneration is considered as the last measure, and not to be carried out regularly.

If the separating properties of the column have deteriorated, the column can be regenerated according to the specifications of the column manufacturer. In the case of separation columns available from Metrohm, the specification for regeneration can be found on the information sheet provided along with each column.

5.16 Quality Management and validation with Metrohm

Quality Management

Metrohm offers you comprehensive support in implementing quality management measures for instruments and software. Further information on this can be found in the brochure **«Quality Management with Metrohm»** available from your local Metrohm agent.

Validation

Please contact your local Metrohm agent for support in validating instruments and software. Here you can also obtain validation documentation to provide help for carrying out the **Installation Qualification** (IQ) and the **Operational Qualification** (OQ). IQ and OQ are also offered as a service by the Metrohm agents. In addition, various application bulletins are also available on the subject, which also contain **Standard Operating Procedures** (SOP) for testing analytical measuring instruments for reproducibility and correctness.

Maintenance

Electronic and mechanical functional groups in Metrohm instruments can and should be checked as part of regular maintenance by specialist personnel from Metrohm. Please ask your local Metrohm agent regarding the precise terms and conditions involved in concluding a corresponding maintenance agreement.



Note

You can find information on the subjects of quality management, validation and maintenance as well as an overview of the documents currently available at www.metrohm.com/com/ under **Support**.

6 Troubleshooting

6.1 Problems and their solutions

Problem	Cause	Remedy
Marked drop in pressure	<i>Leak in the system.</i>	Check and seal connections (see Chapter 3.5, page 19).
	<i>6.2821.120 inline filter blocked.</i>	Replace 6.2821.130 filter (see Chapter 5.6, page 88).
	<i>MSM-HC – blocked.</i>	Regenerate the MSM-HC (see Chapter 5.11.3.1, page 93). Note: 6.2821.180 pump tubing connection with filter must be used .
	<i>Conductivity detector blocked.</i>	- Shorten capillary ends by several mm (see Chapter 5.14.1, Page 102). - Rinse detector against the normal flow direction (see Chapter 5.14.1, Page 102).
	<i>Guard column – blocked.</i>	Replace guard column (see Chapter 3.21, page 67).
	<i>Separation column – blocked.</i>	- Regenerate separation column (see Chapter 5.15.4, page 103). - Replace separation column (see "Connecting and rinsing the separation column", page 70). Note: Samples should always be microfiltered (see Chapter 5.7, page 90).
Drift of the baseline	<i>Injection valve – valve blocked.</i>	Have the valve cleaned (by Metrohm service technicians).
	<i>Thermal equilibrium not yet attained.</i>	Condition (see Chapter 3.15, page 49) instrument with the column thermostat switched on .
	<i>Leak in the system.</i>	Check and seal connections (see Chapter 3.5, page 19).

Problem	Cause	Remedy
	<i>MCS – vacuum pump defective.</i>	Contact the Metrohm Service.
Background conductivity too high	<i>MSM-HC – not connected.</i>	Connecting the MSM-HC (see Chapter 3.16, page 52).
	<i>MCS – not connected.</i>	Connect the MCS.
	<i>Incorrect eluent.</i>	Change eluent (see Chapter 5.4.2.3, page 77).
	<i>MSM-HC – regeneration or rinsing solution flow problems.</i>	Check the flow of the regeneration and rinsing solution (see Chapter 3.16.2, page 52).
Data of the separation column cannot be read.	<i>Column chip contaminated.</i>	Clean contact surfaces of the column chip (with alcohol).
	<i>Column chip defective.</i>	1. Save column configuration in MagIC Net. 2. Notify Metrohm Service.
Individual peaks greater than expected	<i>Sample – Carry-over of the samples from previous measurement.</i>	Rinse system longer between two samples.
MSM-HC – No (or insufficient) pumping of regeneration or rinsing solution	<i>Leak in the system.</i>	Check connections.
	<i>Peristaltic pump – contact pressure too weak.</i>	Correctly set contact pressure .
	<i>Peristaltic pump – filter blocked .</i>	Replace the filter .
	<i>MSM-HC – backpressure too high.</i>	Clean the MSM-HC (see Chapter 5.11.3.2, page 94) or replace parts (see Chapter 5.11.3.3, page 97).
	<i>Peristaltic pump – pump tubing defective.</i>	Replace pump tubing .
Poor reproducibility of the retention times	<i>Eluent – Leak in the eluent path.</i>	Check the eluent path.
	<i>Eluent – Blockage in the eluent path.</i>	Check the eluent path.
Chromatograms have poor resolution	<i>Separation column – diminished separating efficiency.</i>	Regenerate separation column (see Chapter 5.15.4, page 103).

7 Technical specifications

7.1 Reference conditions

The technical data listed in this Chapter refers to the following reference conditions:

Ambient temperature	+25 °C (± 3 °C)
Instrument status	> 40 minutes in operation (equilibrated)

7.2 Instrument

IC system	■ Metal-free IC system ■ Compact system with modular design ■ Up to two complete chromatographic systems in one housing
Material	Painted polyurethane hard foam without CFCs, fire class V0
Operating pressure range	■ 0...50 MPa (500 bar) high pressure pump ■ 0...35 MPa (350 bar) standard-PEEK system
Intelligent components	iPump, iDetector, iColumn, MagIC Net

7.3 Leak sensor

Type	Electronic, no calibration necessary
------	--------------------------------------

7.4 Ambient conditions

<i>Operation</i>	
<i>Ambient temperature</i>	+5...+45 °C
<i>Humidity</i>	20...80 % relative humidity
<i>Storage</i>	
<i>Ambient temperature</i>	-20...+70 °C
<i>Transport</i>	
<i>Ambient temperature</i>	-40...+70 °C

7.5 Housing

Dimensions

Width	365 mm
Height	642 mm
Depth	380 mm

Material of base tray, housing and bottle holder Polyurethane hard foam (PUR) with flame retardation for fire class V0, CFC-free, painted

Operating elements

<i>Indicators</i>	LED for power display
<i>On/Off switch</i>	On the rear panel of the instrument

7.6 Eluent degasser

<i>Material</i>	fluoropolymer
<i>Resistance to solvents</i>	No restriction (apart from PFC)
<i>Build-up time for the vacuum</i>	< 60 s

7.7 High pressure pump

<i>Type</i>	▪ Serial dual-piston pump ▪ Intelligent pump head recognition ▪ Chemically inert ▪ Metal-free pump heads ▪ Materials in contact with eluent: PEEK, ZrO₂, PTFE/PE ▪ Self-optimizing flow and pressure
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Flow rate

<i>Adjustable flow range</i>	0.001...20.0 mL/min
<i>Flow increment</i>	1 µL/min
<i>Reproducibility of the eluent flow</i>	< 0.1 % deviation

Pressure range

<i>Pump</i>	0...50.0 MPa (0...500 bar)
<i>Pump head</i>	0...35.0 MPa (0...350 bar) (applies for the standard PEEK pump head)
<i>Residual pulsa- tion</i>	< 1 %

Safety shutdown

<i>Function</i>	Automatic shutdown upon reaching the pressure limit values
<i>Maximum pressure limit</i>	▪ Adjustable from 0.1...50 MPa (1...500 bar) ▪ The pump is automatically shut down at the first piston stroke above the maximum limit value
<i>Minimum pressure limit</i>	▪ Adjustable from 0...49 MPa (0...490 bar) ▪ The shutdown mechanism is inactive at 0 MPa ▪ The shutdown mechanism only becomes active 2 minutes after system start ▪ The pump is automatically shut down after 3 piston strokes below the minimum pressure limit

<i>Gradient capacity</i>	Isocratic or gradient (extendable to quaternary)
<i>Profile</i>	Step, linear, convex and concave
<i>Resolution</i>	< 1 nL/min flow increments

7.8 Sample degasser

<i>Material</i>	fluoropolymer
<i>Resistance to solvents</i>	No restriction (apart from PFC)
<i>Build-up time for the vacuum</i>	< 60 s

7.9 Injection valve

<i>Actuator time</i>	typ.100 ms
<i>Max. operating pressure</i>	35 MPa (350 bar)
<i>Material</i>	PEEK



7.13 Metrohm CO₂ suppressor (MCS)

<i>Material</i>	fluoropolymer
<i>Resistance to sol- vents</i>	No restriction (apart from PFC)
<i>Vacuum</i>	
<i>Working range</i>	Microprocessor-controlled / stabilized
<i>Build-up time after start</i>	< 30 s
<i>Capillary volume</i>	400 µL
<i>Recommended flow range</i>	0.1 ... 1.0 mL

7.14 Conductivity measuring system

Type	▪ Microprocessor-controlled Digital Signal Processing (DSP technology) ▪ Intelligent detector with 6 reference chromatograms
Measuring range	0...15000 $\mu\text{S}/\text{cm}$ single range
Noise	< 0.1 nS at 1 $\mu\text{S}/\text{cm}$
Deviations from the linearity	< 1 % for conductivity values greater than 1...16 $\mu\text{S}/\text{cm}$ (typical for analyses with sequential suppression)
Drift	< 0.2 nS/cm per hour
Measuring rate	10 measurements per second for optimum results without filtering
Resolution	0.0047 nS/cm
Baseline	Noise < 0.2 nS/cm typical for sequential suppression
Conductivity detector	
Cell volume	0.8 μL
Cell constant	▪ Individual calibration data saved in the detector ▪ adjustable range: 13.0...21.0 /cm
Electrodes	Ring-shaped electrodes made from stainless steel
Materials in contact with eluent	Chemically inert PCTFE
Maximum operating pressure	5.0 MPa (50 bar)

<i>Cell temperature</i>	20...50 °C in increments of 5 °C
<i>Temperature stability</i>	< 0.001 °C
<i>Temperature compensation</i>	0...5 %/K adjustable, default 2.3 %/K
<i>Heating up time</i>	< 30 minutes (40 °C)

7.15 Mains connection

<i>Required voltage</i>	100...240 V $\pm$ 10 % (auto-sensing)
<i>Required frequency</i>	50...60 Hz $\pm$ 3 (auto-sensing)
<i>Power consumption</i>	65 W for typical analysis application 25 W standby (detector to 40 °C)
<i>Power supply unit</i>	- Up to 300 W maximum, electronically monitored - internal fuse 3.15 A

7.16 Interfaces

USB

<i>Input</i>	1 USB upstream, type B (for connection to the PC)
<i>Output</i>	2 USB downstream, type A

<i>MSB</i>	2 MSB Mini Din 8-pin (female) (for Dosino, stirrer, remote lines, ...)
------------	--



Caution

When connecting an instrument to the MSB connector you **must** switch off the 850 Professional IC .

<i>Detector</i>	2 DSUB-15-pin high density (female)
<i>Column recognition</i>	3 (including 2 in the column thermostat (see Chapter 3.15, page 49))
<i>Leak sensor</i>	1 jack plug
<i>Further connections</i>	1 DSUB 15-pin (female)



7.17 Safety specification

Design / Test

- EN/IEC/UL 61010-1
- CSA-C22.2 No. 61010-1
- Degree of protection IP20
- Protection class I

7.18 Electromagnetic compatibility (EMC)

Emission

- EN/IEC 61326-1
- EEN 55022 / CISPR 22
- EN/IEC 61000-6-3
- EN/IEC 61000-3-2
- EN/IEC 61000-3-3

Immunity

- EN/IEC 61326-1
- EN/IEC 61000-4-2
- EN/IEC 61000-4-3
- EN/IEC 61000-4-4
- EN/IEC 61000-4-5
- EN/IEC 61000-4-6
- EN/IEC 61000-4-8
- EN/IEC 61000-4-11
- EN/IEC 61000-4-14
- NAMUR: 2006

7.19 Weight

1.850.2220	33.1 kg (without accessories)
1.850.9010 (conductivity detector)	2.3 kg (with accessories)
Transport carriage (rollers and handle)	1.8 kg



FN 61010-1

Safety requirements for electrical equipment for measurement, control and laboratory use



This instrument meets the requirements of the ETL Listed Mark for the North American market. It conforms to the electrical safety standards UL 61010-1 and CSA-C22.2 No. 61010-1. This product is listed in Intertek's Directory of Listed Products.

Manufacturer

Metrohm Ltd., CH-9101 Herisau/Switzerland

Metrohm Ltd. is holder of the SQS-certificate ISO 9001:2000 Quality management system for development, production and sales of instruments and accessories for ion analysis.

Herisau, 27 October, 2008

D. Schum

D. Strohm

Vice President, Head of R&D

A. B. C. D. E. F. G. H. I. J. K. L. M. N. O. P. Q. R. S. T. U. V. W. X. Y. Z.

Ch. Buchmann

Vice President, Head of Production

Responsible for Quality Assurance

8.2 Quality Management Principles

Metrohm Ltd. holds the ISO 9001:2000 Certificate, registration number 10872-02, issued by SQS (Swiss Association for Quality and Management Systems). Internal and external audits are carried out periodically to assure that the standards defined by Metrohm's QM Manual are maintained.

The steps involved in the design, manufacture and servicing of instruments are fully documented and the resulting reports are archived for ten years. The development of software for PCs and instruments is also duly documented and the documents and source codes are archived. Both remain the possession of Metrohm. A non-disclosure agreement may be asked to be provided by those requiring access to them.

The implementation of the ISO 9001:2000 quality management system is described in Metrohm's QM Manual, which comprises detailed instructions on the following fields of activity:

Instrument development

The organization of the instrument design, its planning and the intermediate controls are fully documented and traceable. Laboratory testing accompanies all phases of instrument development.

Software development

Software development occurs in terms of the software life cycle. Tests are performed to detect programming errors and to assess the program's functionality in a laboratory environment.

Components

All components used in the Metrohm instruments have to satisfy the quality standards that are defined and implemented for our products. Suppliers of components are audited by Metrohm as the need arises.

Manufacture

The measures put into practice in the production of our instruments guarantee a constant quality standard. Production planning and manufacturing procedures, maintenance of production means and testing of components, intermediate and finished products are prescribed.

Customer support and service

Customer support involves all phases of instrument acquisition and use by the customer, i.e. consulting to define the adequate equipment for the analytical problem at hand, delivery of the equipment, user manuals, training, after-sales service and processing of customer complaints. The Metrohm service organization is equipped to support customers in implementing standards such as GLP, GMP, ISO 900X, in performing Operational Qualification and Performance Verification of the system components or in carrying out the System Validation for the quantitative determination of a substance in a given matrix.

8.3 Warranty (guarantee)

Metrohm guarantees that the deliveries and services it provides are free from material, design or manufacturing errors. The warranty period is 36 months from the day of delivery; for day and night operation it is 18 months. The warranty remains valid on condition that the service is provided by an authorized Metrohm service organization.

Glass breakage is excluded from the warranty for electrodes and other glassware. The warranty for the accuracy corresponds to the technical specifications given in this manual. For components from third parties that make up a considerable part of our instrument, the manufacturer's warranty provisions apply. Warranty claims cannot be pursued if the Customer has not complied with the obligations to make payment on time.

During the warranty period Metrohm undertakes, at its own choice, to either repair at its own premises, free of charge, any instruments that can be shown to be faulty or to replace them. Transport costs are to the Customer's account.



Faults arising from circumstances that are not the responsibility of Metrohm, such as improper storage or improper use, etc. are expressly excluded from the warranty.

9 Accessories



Subject to change without notice!

9.1 Scope of delivery

2.850.2240 850 Professional IC – Anion – MSM-HC – MCS – Gradient

Qty.	Order no.	Description
1	1.850.2240	850 Professional IC Anion – MSM-HC – MCS – Gradient
1	6.2122.0x0	Mains cable with C13 line socket IEC-60320-C13
		Cable plug according to customer requirements.
	Switzerland:	Type SEV 12 6.2122.020
	Germany, ...:	Type CEE(7), VII 6.2122.040
	USA, ...:	Type NEMA/ASA 6.2122.070
1	1.850.9010	IC Detector MF
2	6.1602.150	Bottle neck attachment / GL 45 - 3 x UNF 10/32
		For connecting capillary tubing 1/16 in. (=1.6 mm). Used for MSM auxiliary solutions and in inline dialysis.
	Material:	Plastic
2	6.1602.160	Eluent bottle attachment GL 45
		For eluent bottles; with connections for drying tube and aspiration tubing.
	Opening ground joint:	A-14/15






Qty.	Order no.	Description	
2	6.1608.020	Glass bottle / 1000 mL / GL 45	
		Bottle for auxiliary solutions	
		Width (mm):	
		Height (mm):	
		Volume (mL):	1000
2	6.1608.070	Eluent bottle / 2 L / GL 45	
		Material:	Clear glass
		Height (mm):	262
		Volume (mL):	2000
2	6.1609.000	Adsorbing tube, large and bent	
		For filling with adsorber material.	
		Material:	Glass
		Height (mm):	129
		Inner diameter (mm):	32
		SGJ size:	B-14/15
1	6.1803.020	PTFE capillary 0.97 mm i.d. / 5 m	
		For all IC instruments	
		Material:	PTFE
		Outer diameter (mm):	1.57
		Inner diameter (mm):	0.97
		Length (m):	5





Qty.	Order no.	Description	
2	6.1816.020	Silicone tubing 6 mm i.d., 1 m	
		For waste tubing. Material: Silicone rubber Outer diameter (mm): 9 Inner diameter (mm): 6 Length (m): 1	
2	6.1826.320	Pump tubing LFL (orange/yellow), 3 stoppers	
		For suppressor solutions, acceptor solution in Inline Dialysis and for Inline Ultrafiltration	
2	6.2023.020	Clip for SGJ 14/15	
		Clip for SGJ 14/16 Material: POM	
1	6.2057.080	Adsorption cartridge holder (ProfIC)	
		Holder to install adsorption cartridges in Professional IC Instruments	





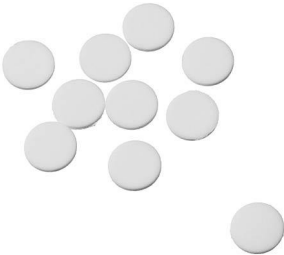
Qty.	Order no.	Description	
1	6.2621.050	1/4 in. wrench For 1/4 in. screws. For IC instruments Length (mm): 73	
1	6.2621.080	Capillary cutter For plastic capillaries. Used with IC instruments Length (mm): 118	
1	6.2621.100	Hexagon key 3 mm Hexagon key 3 mm for IC Sample Processors Length (mm): 73	









Qty.	Order no.	Description	
2	6.2816.020	Syringe 10 mL with Luer connection	
		For various applications in IC and VA	
		Material:	
		Length (mm):	
		Volume (mL):	
1	6.2816.040	Purging needle	
		With PTFE tubing and Luer connection. For syringes. For aspirating eluents.	
2	6.2821.090	Aspiration filter	
		Pore size 20 µm, set of 5 pieces. For 6.1834.000 Aspiration tubing and 6.1821.040 and 6.1821.050 Filter tubes.	
		Material:	
		Outer diameter (mm):	
		Length (mm):	
2	6.2821.130	Spare filter for inline filter	
		Spare filters for inline filter.	

Qty.	Order no.	Description	
1	6.2837.000	CO₂ adsorber cartridge Adsorber cartridge for purifying the air.	
2	6.2837.010	H₂O adsorption cartridge To CO ₂ Suppressor. Water adsorption cartridge for the aspirated air.	
1	8.850.8027EN	Manual for 850 Professional IC, 2.850.2240 – Anion – MSM-HC – MCS – Gradient, English	

2.850.2240 850 Professional IC – Anion – MSM-HC – MCS – Gradient

Order no.	Description
6.2617.040	Tool for piston seal, macro For removing and assembling the piston seal for all macro pump heads
6.2741.040	PE/PTFE piston seal, macro For all macro pump heads

6.2824.130 Macro pump head PEEK

Macro pump head for intelligente IC instruments, flow range 0.1...20 mL/min, maximum pressure 12.5 MPa.

Material: PEEK (metal-free)



6.6059.112	MagIC Net™ 1.1 Professional CD: 1 license
------------	---

Professional PC program for controlling intelligent Professional IC systems, Compact IC instruments and their peripherals such as Professional Sample Processors, 800 Dosino, 771 Compact Interface, etc. The software permits control, data acquisition, evaluation and monitoring as well as report generation for ion chromatographic analyses. MagIC Net™ comprises the following features: graphic user interface for routine operations, extensive database programs, method development, configuration and manual system control, highly flexible user administration, efficient database operations, extensive data export functions, individually configurable report generator, control and monitoring of all system components and the chromatographic results. MagIC Net™ complies with FDA regulation 21 CFR part 11 as well as GLP. Dialog languages: German, English, French, Chinese, Korean, Japanese and more. 1 license.



6.6059.113	MagIC Net™ 1.1 Multi CD: 3 licenses
------------	-------------------------------------

Professional PC program for controlling intelligent Professional IC systems, Compact IC instruments and their peripherals such as Professional Sample Processors, 800 Dosino, 771 Compact Interface, etc. The software permits control, data acquisition, evaluation and monitoring as well as report generation for ion chromatographic analyses. MagIC Net™ comprises the following features: graphic user interface for routine operations, extensive database programs, method development, configuration and manual system control; highly flexible user administration, efficient database operations, extensive data export functions, individually configurable report generator, control and monitoring of all system components and the chromatographic results. MagIC Net™ complies with FDA regulation 21 CFR part 11 as well as GLP. Dialog languages: German, English,



Quality Management with Metrohm

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Ion analysis

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