



Application Note AN-NIR-093

# Quality Control of fermentation processes

## Multiparameter determination within one minute

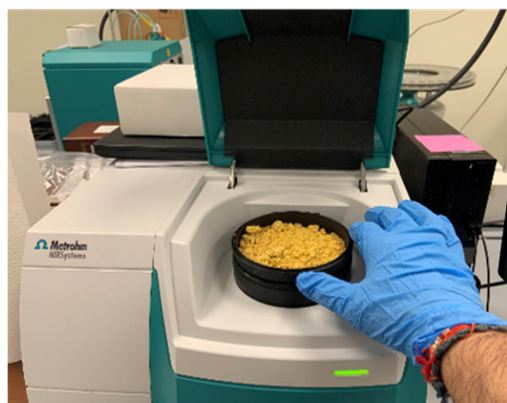
The production of biofuels from renewable feedstock has grown immensely in the past several years. Bioethanol is one of the most interesting alternatives for fossil fuels, since it can be produced from raw materials rich in sugars and starch. Ethanol fermentation is one of the oldest and most important fermentation processes used in the biotechnology industry. Although the process is well-known, there is a great potential for its improvement and a

proportional reduction in production costs. Due to the seasonal variation of feedstock quality, ethanol producers need to monitor the fermentation process to ensure the same quality product is achieved.

Near-infrared spectroscopy (NIRS) offers **rapid and reliable prediction** of ethanol content, sugars, Brix, lactic acid, pH, and total solids at any stage of the fermentation process.

## EXPERIMENTAL EQUIPMENT

Production of ethanol from corn goes through three typical steps: milling / liquefaction of corn into starch mash, fermentation of starch mash with yeast, and finally purification of the resulting ethanol by distillation. A total of 206 samples (117 for Brix index) of fermentation mash were analyzed on the DS2500 Solid Analyzer. Due to the large amount of solids present in the samples, all measurements were performed in reflection mode using the DS2500 Large sample cup (Figure 1). The samples were measured in rotation to collect spectral data from several areas. Spectral averaging of signals from several spots helped to reduce sample inhomogeneity. The Metrohm software package Vision Air Complete was used for all data acquisition and prediction model development.



**Figure 1.** Fermentation mash sample placed on the DS2500 Solid Analyzer.

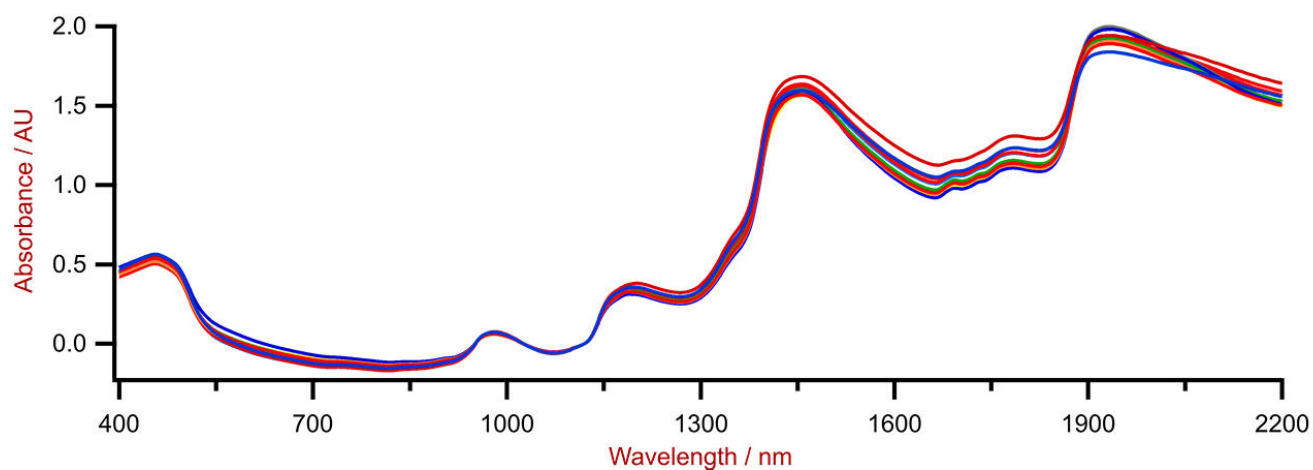
**Table 1.** Hardware and software equipment overview

| Equipment               | Metrohm number |
|-------------------------|----------------|
| DS2500 Analyzer         | 2.922.0010     |
| DS2500 Large Sample Cup | 6.7402.050     |
| Vision Air 2.0 Complete | 6.6072.208     |

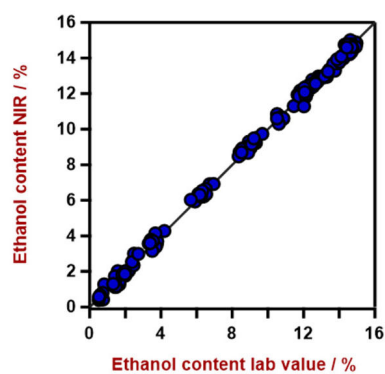
## RESULT

All 206 measured Vis-NIR spectra (Figure 2) were used to create a prediction model for quantification of the key fermentation parameters. The quality of the prediction model was evaluated using correlation diagrams, which display a very high correlation

between the Vis-NIR prediction and the reference values. The respective figures of merit (FOM) display the expected precision of a prediction during routine analysis.



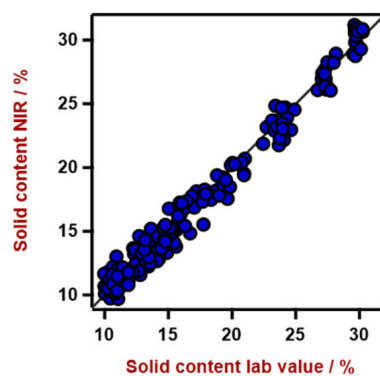
**Figure 2.** Vis-NIR spectra of fermentation mash samples analyzed on a DS2500 Solid Analyzer.



**Figure 3.** Correlation diagram for the prediction of ethanol content using a DS2500 Solid Analyzer. The ethanol content lab value was evaluated using HPLC.

**Table 2.** Figures of merit for the prediction of ethanol content using a DS2500 Solid Analyzer.

| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.998 |
| Standard error of calibration      | 0.21% |
| Standard error of cross-validation | 0.22% |

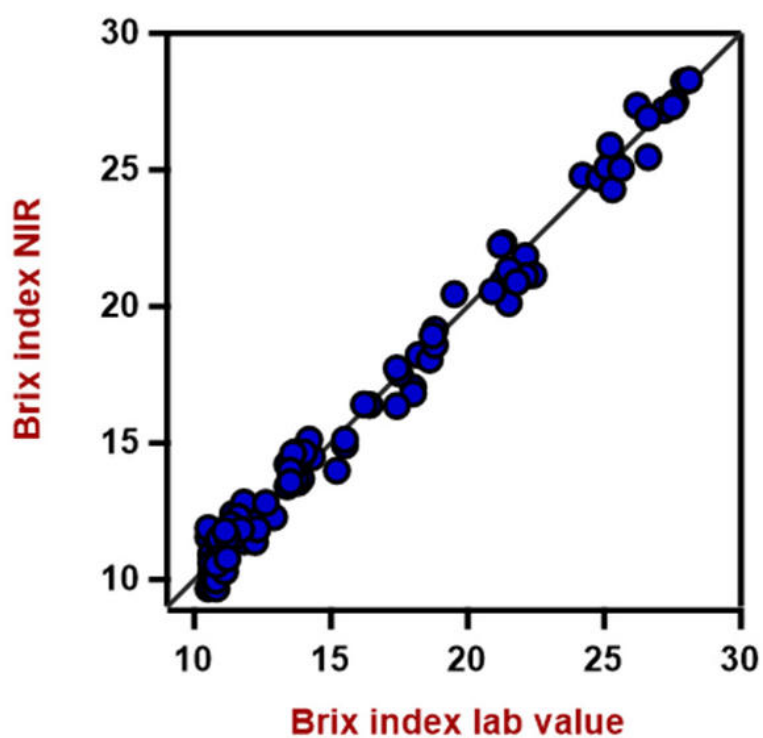


**Figure 4.** Correlation diagram for the prediction solid content using a DS2500 Solid Analyzer. The lab value was evaluated by LOD balance.

**Table 3.** Figures of merit for the prediction solid content using a DS2500 Solid Analyzer.

| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.982 |
| Standard error of calibration      | 0.87% |
| Standard error of cross-validation | 1.06% |

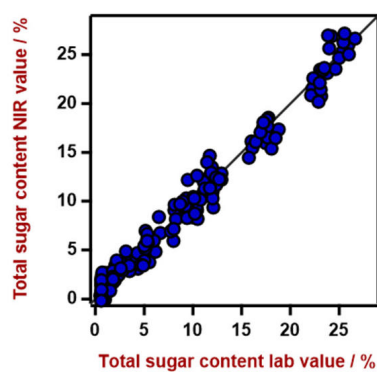
## RESULT



**Figure 5.** Correlation diagram for the prediction of Brix index values. The lab value was measured using a refractometer.

**Table 4.** Figures of merit for the prediction of Brix index values.

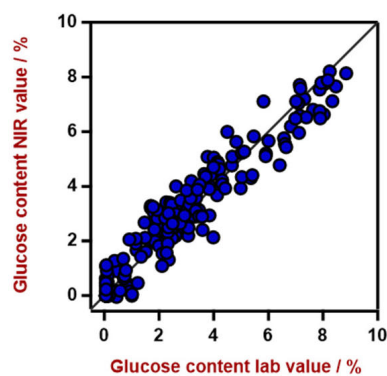
| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.987 |
| Standard error of calibration      | 0.66  |
| Standard error of cross-validation | 0.87  |



**Figure 6.** Correlation diagram for the prediction of the total sugar content. The total sugar content lab value was measured using HPLC.

**Table 5.** Figures of merit for the prediction of the total sugar content.

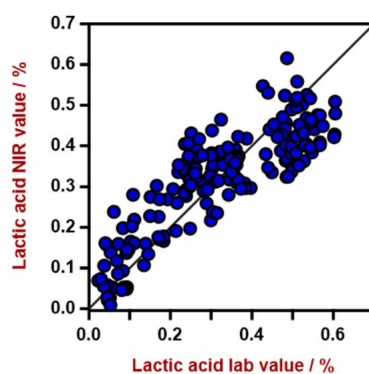
| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.981 |
| Standard error of calibration      | 1.09% |
| Standard error of cross-validation | 1.30% |



**Figure 7.** Correlation diagram for the prediction of glucose content. The glucose content lab value was measured using HPLC.

**Table 6.** Figures of merit for the prediction of the glucose content.

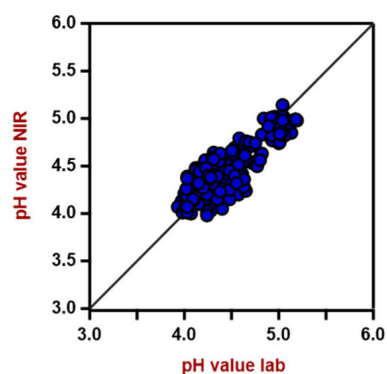
| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.920 |
| Standard error of calibration      | 0.70% |
| Standard error of cross-validation | 0.86% |



**Figure 8.** Correlation diagram for the prediction of lactic acid content. The lactic acid lab value was evaluated using HPLC.

**Table 7.** Figures of merit for the prediction of lactic acid content.

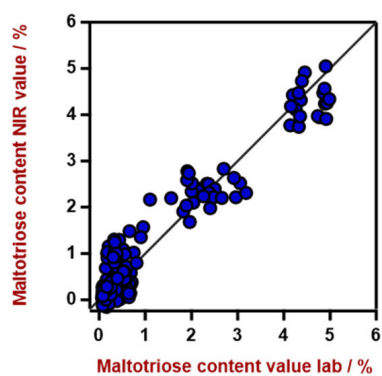
| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.722 |
| Standard error of calibration      | 0.09% |
| Standard error of cross-validation | 0.10% |



**Figure 9.** Correlation diagram for the prediction of pH value. The pH lab value was measured using a pH meter.

**Table 8.** Figures of merit for the prediction of pH value.

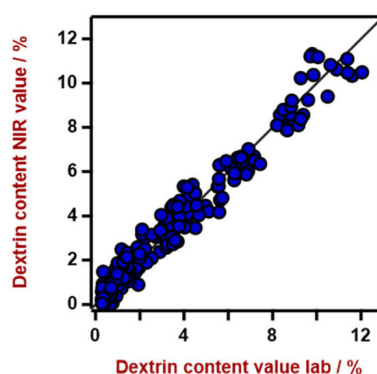
| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.734 |
| Standard error of calibration      | 0.17  |
| Standard error of cross-validation | 0.19  |



**Figure 10.** Correlation diagram for the prediction of maltotriose content. The maltotriose lab value was measured using HPLC.

**Table 9.** Figures of merit for the prediction of maltotriose content.

| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.928 |
| Standard error of calibration      | 0.36% |
| Standard error of cross-validation | 0.42% |



**Figure 11.** Correlation diagram for the prediction of dextrin content. The dextrin lab value was measured using HPLC.

**Table 10.** Figures of merit for the prediction of dextrin content.

| Figures of merit                   | Value |
|------------------------------------|-------|
| $R^2$                              | 0.964 |
| Standard error of calibration      | 0.60% |
| Standard error of cross-validation | 0.68% |

## CONCLUSION

This application note demonstrates the feasibility to determine multiple key parameters of the fermentation process with NIR spectroscopy. Corn fermentation is a well-established process which typically runs for 55–60 hours. Samples are extracted from fermenters every few hours and sent to the

laboratory for analytical measurement. Several analytical methods need to be used to monitor key quality parameters for the fermentation process. Vis-NIR spectroscopy enables a fast alternative with high accuracy, and therefore represents a suitable single method to monitor the fermentation process.

**Table 11.** Time to result overview for the different parameters

| Parameter       | Method        | Time to result |
|-----------------|---------------|----------------|
| Ethanol, sugars | HPLC          | 30–45 min      |
| Brix index      | Refractometer | 3–5 min        |
| pH              | pH meter      | 3–5 min        |
| Solids          | LOD Balance   | 10–15 min      |

## CONTACT

Metrohm Middle East FZC  
E2-05 SAIF Zone  
120747 Sharjah, UAE

[info@metrohm.ae](mailto:info@metrohm.ae)

## EXPERIMENTAL EQUIPMENT



### DS2500 Solid Analyzer

Robust near-infrared spectroscopy for quality control, not only in laboratories but also in production environments.

The DS2500 Analyzer is the tried and tested, flexible solution for routine analysis of solids, creams, and optionally also liquids along the entire production chain. Its robust design makes the DS2500 Analyzer resistant to dust, moisture, vibrations, and temperature fluctuations, which means that it is eminently suited for use in harsh production environments.

The DS2500 covers the full spectral range from 400 to 2500 nm and delivers accurate, reproducible results in less than one minute. The DS2500 Analyzer meets the demands of the pharmaceutical industry and supports users in their day-to-day routine tasks thanks to its simple operation.

Thanks to accessories tailored perfectly to the instrument, optimum results are achieved with every sample type, no matter how challenging it is, e.g. coarse-grained solids such as granulates or semi-solid samples such as creams. The MultiSample Cup can help improve productivity when measuring solids, as it enables automated measurements of series containing up to 9 samples.



### DS2500 large sample cup

Large sample cup for the spectral recording of powders and granulates in reflection at various sample positions using the NIRS DS2500 Analyzer.



## Vision Air 2.0 Complete

Vision Air - Universal spectroscopy software.

Vision Air Complete is a modern and simple-to-operate software solution for use in a regulated environment.

Overview of the advantages of Vision Air:

- Individual software applications with adapted user interfaces ensure intuitive and simple operation
- Simple creation and maintenance of operating procedures
- SQL database for secure and simple data management

The Vision Air Complete version (66072208) includes all applications for quality assurance using Vis-NIR spectroscopy:

- Application for instrument and data management
- Application for method development
- Application for routine analysis

Additional Vision Air Complete solutions:

- 66072207 (Vision Air Network Complete)
- 66072209 (Vision Air Pharma Complete)
- 66072210 (Vision Air Pharma Network Complete)